Online ISSN 2559 - 1061

INTERACTION OF BIOACTIVE COMPOUNDS WITH CERAMIC MATERIALS – A REVIEW

Gertrud-Alexandra PALTINEAN¹, Gheorghe TOMOAIA^{2,3}, Levente-Zsolt RACZ¹, Aurora MOCANU¹, Maria TOMOAIA-COTISEL^{1,3}

Abstract. This review examines the interaction between silymarin (SIL) and other plantbased bioactive compounds such as curcumin (CCM), piperine (PIP), resveratrol (RES), and icariin (ICA). Their combination revealed synergistic effects on colon (HCT116), breast (T47D) tumor cells, hepatocellular carcinoma, and periodontal disease. The review also addresses the interaction between these plant extracts with ceramic materials such as hydroxyapatite (HAP) and carotenoids with concrete examples of biomedical applications. Silymarin's interaction with chemotherapeutic drugs (doxorubicin-DOX, paclitaxel-PCT and 5-Fluorouracil-5-FLU) and gold nanoparticles-GNPs and silver nanoparticles-SNPs is also debated. All these combinations can form composites of major importance in the biomedical field and to contribute significantly to orthopedic surgery where materials are needed for implants that face severe infections. This short review highlights the variety of multifunctional nanoparticles that open new opportunities in cancer treatment and the need to use the Langmuir Blodgett Technique that mimics the biological membrane and provides rich medical information.

Keywords: Silymarin, bioactive compounds, hydroxyapatite, carotenoids, chemotherapeutic drugs, gold nanoparticles, silver nanoparticles

https://doi.org/10.56082/annalsarsciphyschem.2022.2.47

1. Introduction

Plants and their role in human health have recently received special attention. They have in their portfolio a wide range of biological activities with the role of protecting the organs and tissues of the body against various diseases. By using them, it has been possible to maintain a balance between controlling morbidity and regaining health. The medical system is exposed to pathogens. This means maintaining health is difficult for those hospitalized for either treatment or chirurgical procedures. Science through its research activities and experimental results highlights a first step in prolonging health by preventing disease and efficacy against widespread communicable diseases. In this sense, some bioactive compounds, seen in figure 1, fight against diseases since ancient times.

³Academy of Romanian Scientist, 3 Ilfov Str., District 5, RO 050044, Bucharest, Romania *Corresponding author: Maria Tomoaia-Cotisel, <u>mcotisel@gmail.com</u>

¹Babes-Bolyai University, Faculty of Chemistry and Chemical Engineering, Research Centre of Physical Chemistry, 11 Arany Janos Str., RO 400028, Cluj-Napoca, Romania

²Iuliu Hatieganu University of Medicine and Pharmacy, Department of Orthopedic Surgery, 47 General Traian Mosoiu Str., RO 400132, Cluj-Napoca, Romania

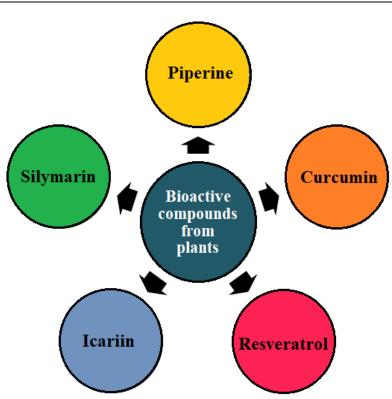


Fig. 1. Bioactive compounds from plants

The previous study carried out by our team showed a rich biological activity and therapeutic potentials of piperine, curcumin, resveratrol, and icariin. Also, we discussed the interaction between them, the interaction with chemotherapeutic drugs, the interaction with silver and gold nanoparticles and the beneficial effects on health, especially applications in cancer cell lines [1].

In the present study, all these proactive compounds and silymarin have in common their anticancer and anti-inflammatory activities. However, the most studied at present is the anticancer activity. Cancer is a relentless disease that does not consider age and involves the abnormal development of body cells. It can be established in any type of organ such as pancreas, liver, lungs, breast, ovaries, stomach, rectum, etc. This disease can occur either inherited or due to multiple environmental factors and socio-industrial activities in which man lives. Examples that can lead to cancer are smoking, alcohol, poor lifestyle, unhealthy diet, exposure to toxic substances. From this point of view, these compounds could become the main pawns in the treatment of this condition and, why not, the definitive eradication in a stage in which the disease is not advanced enough so that the tissues respond effectively to the treatment.

Curcumin (CCM) is a bioactive compound produced by *Curcuma Longa* and is currently used as a dye in the food industry. The solubility of curcumin is poor and the rate of absorption into the body is also low. Due to its rich biological activities, it has been used since ancient times in traditional Chinese, Indian and Ayurvedic medicine and is widely used in biomedical applications. In the case of anti-cancer activity curcumin downregulates the production of pro-inflammatory cytokines TNF- α and IL-1 β and inhibits the activation of transcription factors nuclear NF-kB [2]. Also, it is known to induce apoptosis in prostate cancer cells but also to prevent the progression of this cancer [3]. This proactive compound can form innovative composites and complexes with applications in medicine and in this sense facilitates the understanding of the mechanisms of action during application [4-9].

Silymarin (SIL) is the main component in the plant Silybum marianum, also used since ancient times in the treatment of liver diseases (cirrhosis, jaundice, hepatitis) and biliary diseases [10-12]. It is a compound rapidly metabolized, absorbed and eliminated within six hours. It is not soluble in water but can be administered as an encapsulated standardized extract [13, 14]. SIL has a rich biological activity such as anticancer, antioxidant, anti-inflammatory, antidiabetic, anti-lipemic, anti-osteoporotic, anti-viral, anti-arthritic [11, 13]. It is widely studied in the treatment of cancer because it inhibits tumor growth and modulates signaling pathways such as NF- β , EGFR-MAPK / ERK 1/2 and IGF. In the case of bladder carcinoma, silymarin stops the G2 / M phase and modulates the cascade of the cyclin CDK1-CDK. It is reported to activate caspase 3, resulting in inhibition of the growth and apoptotic death of TCC cells [11, 15]. In hepatocellular carcinoma, silymarin inhibits the increase in b-catenin, inhibits mitochondrial membrane potential of HepG2 cells and modulates the activity of CDK-2, CDK-4, and CDC-2 kinase [16]. In prostate cancer silymarin inhibits the growth of cancer cells both in vitro and in vivo and modulates MAPK, ERK 1/2, and IGF signaling pathways [17]. In the case of A549 cells from lung cancer, silymarin inhibits phosphorylation of ERK 1/2 and reduces the level of MMP-2 and u-PA [18].

It is known from literature studies that **piperine** (**PIP**) inhibits the growth of human prostate cancer cells such as LNCaP, PC-3 and DU145 in a dosedependent manner and induces the cell cycle arrest at G0/G1-phase (P < 0.05). It also reduces the viability of osteosarcoma cells (HOS, U2OS) in time and dosedependent manners and exposure to piperine causes G2/M phase arrest of the cell cycle. On the human bone marrow (K-562 leukemic cells) piperine induces anticancer effects. The fight against tumor cancer lines is in a dose-dependent manner, in which the mechanism of action is associated with mitochondrial damage, increased reactive oxygen species and expression of key proteins (Bcl - 2, Bax, Cyt - c, Caspase - 9 and Caspase - 3) affected [19-22].

In the case of bladder cancer, **resveratrol** (**RES**) exerts a significant cytotoxic effect and induced cellular apoptosis of T24 cells in a dose- and time-dependent manner. Also, the treatment of T24 cells with resveratrol caused G1 phase cell cycle arrest [23]. Resveratrol has low bioavailability and is known to act synergistically with hemotherapeutic drugs to increase their anti-cancer effects. It was demonstrated that resveratrol sensitized chemotherapeutic drugs resistant to cancers [24, 25]. Although the mechanism of action of resveratrol is not fully elucidated it has been shown that it directly inhibits the proliferation of pancreatic cancer cells in a dose- and time-dependent manner. Also, it can induce apoptosis and cell cycle arrest, enhances the chemo-radio-sensitization and can affect diabetes mellitus [26].

As for **icariin** (**ICA**), it has anticancer activity in lung cancer. In vivo experiments have shown a decrease in H1975 cell proliferation and in vitro this reduces A549 and H1975 cell proliferation in a dose and time dependent manner [27]. In the case of human esophageal carcinoma cells (KYSE70 cells), icariin causes the cell cycle to stop in the G2 / M phase [28]. Icariin, a hydrolytic product of icariin, has also been widely studied in traditional Chinese medicine. Nowadays, due to the multiple pharmacological activities, icariin has increased the interest of researchers for antitumor activity. In the case of different types of cancer such as human endometrial cancer and glioblastoma multiform cell lines, icariin induces G1 phase arrest. In lung cancer, it causes S phase arrest while in oral squamous cell carcinoma (OSCC), colon cancer and renal cell carcinoma cell lines; it arrests cell cycle in G1/S phase [29].

Given that technology and nanoscience have reached a high level of development, the World Health Organization (WHO) estimates that about a third of all existing cancers could be prevented [30, 31]. According to the WHO globally, in 2018, 18.1 million people were diagnosed with cancer, of which 9.6 million died. The most common cancers are lung (11.6%), breast (11.6) and colorectal (10.2%). Early diagnosis is the best way to prevent the progression of this disease followed by treatment that may involve chemotherapy and surgery. In the case of cancer that occurs in children, the most common is bone marrow leukemia and lymphatic system cancer. The cause of the emergence is not yet understood, but at the level of current knowledge it is known that it is not preventable.

Thus far, we have highlighted the characteristics of the bioactive compounds applied individually; in the following we will discuss the synergistic effect obtained between Silymarin and these compounds. This herbal therapy enhances beneficial activity in treating diseases and involves mechanisms of action responsible for stopping cancer at various stages of development.

2. Silymarin interaction with proactive compounds from plants

Literature studies show that there is a synergistic effect between the *silymarin* and curcumin. A. Montgomery and colleagues studied the effect of this combination using colon cancer cell lines DLD-1, HCT116 and LoVo. They showed that the combined treatment on these cancerous lines inhibited cell proliferation and increased apoptosis. As a conclusion of their study, the combination of CCM + SIL leads to higher level of inhibition of cancer cell much more effectively than if applied individually [32].

A study conducted by N. Abdel-Magied highlights the possible curative role of combined therapy against nephrotoxicity induced by gamma-rays in rats. The irradiated rats treated with combined therapy revealed a significant decrease of malondialdehyde, H₂O₂ and advanced oxidation protein products and a significant increase of GSH and total antioxidant capacity. They observed additive effects on the level of Interleukin 18, tumor necrosis factor alpha, C-reactive protein, Bax, factor-related apoptosis and the activity of Casp-3 by 58%, 58%, 41%, 47%, 64%, 31%, respectively associated with an increase of Bcl2 level by 122%. The additive effect is manifested by a decrease of malondialdehyde, hydrogen peroxide and advanced oxidation protein products on irradiated rats. They conclude that combination could be used as a medication for protection of patients during radiotherapy [33].

M. M. Ahmad et al show that paracetamol treatment caused liver damage. Individual application of CCM and SIL produced hepatoprotective effects due to antioxidant activities. They conclude that CCM is more effective than SIL at protecting the liver against paracetamol toxicity [34]. T47D breast cancer cell line was treated with Silibinin+CCM mixture. This mixture has an inhibitory effect on hTERT gene expression in a dose-dependent manner and the real-time PCR results show a significant decrease in hTERT expression [35].

Silymarin and resveratrol combination was highlighted in a study conducted by A. Farzanegan and coworkers. The obtained results show antihistamine effects on human gingival fibroblasts. They evaluate the cell viability in 24h and 48h after treatment with 50/100 and 100/200 µg/ml of silymarin/resveratrol in the presence of histamine (10 µg/ml). The results showed a significant reduction in viability with a combination of 100 µg/ml silymarin and 200 µg/ml resveratrol in 48 h. This concentration was found to decline the secretion of IL-8 and TPA-1 while the combination of resveratrol/silymarin 100 µg/ml and 50 µg/ml demonstrated a considerable drop in the secretion of all detected factors (IL-8, IL-6, TPA-1, and TNF- α). This kind of combination can be useful as a therapeutic agent for treatment of periodontal diseases [36, 37]. In their study, W.-C. Hsieh et al concluded by their study that the Silymarin/resveratrol combination presents a synergistic effect on the reduction of the hepatocellular carcinoma in a mouse model and can be a potential agent for the prevention of HCC in high-risk chronic hepatitis B virus carriers [38].

Regarding the combination of *piperine and silymarin*, R. Shukla and collaborators, evaluate the hepatoprotective activity against D-galactosamine induced liver damage in albino rats. The results after treatment with this combination show a protective activity on liver against the injury induced by D-galactosamine [39]. S. Javed et al suggest by their studies that SIL + PIP might have a synergistic effect and might have hepatoprotective and antioxidant activity [40].

Regarding ICA and SIL, there is not enough information to evaluate the action mechanism.

3. Silymarin and chemotherapeutic drugs

In our previous study we highlighted the interaction of plant extracts with chemotherapeutic drugs (doxorubicin - DOX, paclitaxel - PCT and 5fluorouracil-5-FLU) and separately with gold and silver nanoparticles. Silvmarin can also interact with chemotherapeutic drugs to help treat different types of cancer and other conditions. Studies in the literature have investigated the genotoxic and cytotoxic effects of SIL and DOX applied individually and in combination on the HepG2 cell line for 24 hours and 48 hours. E. Yurtcu et al noticed that the individual application of the two is more effective than their combination in this case. They say that after 24 hours the two treatments caused DNA damage, while after 48 hours the genotoxic damage was stronger with doxorubicin than with SIL. The combination of the two did not have significant results but more clinical trials are needed to see the effect on other cancer [41]. Another study conducted by N. Patel et al show that SIL reduced DOX hepatotoxicity and associated apoptotic and necrotic cell death. Also, SIL can modulate changes in Bcl-xL and p53 expression [42]. It is known that doxorubicin can cause heart, liver, and kidney toxicity if it is administered more than necessary. E Cecen and his research group demonstrate that silymarin protected these organs from doxorubicin toxicity [43]. F. Gheybi and coworkers combined the SIL + DOX liposomes in 4T1 breast cancer cells at 100 and 300 molar ratios of the two drugs. The results show synergistic growth-inhibitory effects at this molar ratio. They concluded that the successful combination of the two medications plays an important role which determines the final response following treatment [44]. The principal compound from silymarin is silibinin and A. K. Tyagi demonstrated that silibinin strongly synergized the growth-inhibitory effect of doxorubicin in prostate carcinoma DU145 cells with a strong G2-M

phase arrest in cell cycle progression [45]. In the case of **paclitaxel** (**PCT**), another chemotherapeutic drug, silibinin enhanced the therapeutic potential of paclitaxel against human gastric cancer SGC-7901 cells [46]. O. Molavi et al, evaluate the synergistic effect of silibinin and DOX and *silibinin with PCT* to in breast cancer cells line (MDA-MB435 and MCF-7) and the results show better effects of silibinin - DOX in MDA-MB435/WT cells then in silibinin + PCT in MCF-7/WT. They observed that silibinin increase the cytotoxic effect of DOX and PCT [47].

S. Patel et al evaluated the synergy effect of *Silibinin and 5-Fluorouracil* that inhibited cell proliferation of CD44+ subpopulation of human colon carcinoma (HCT 116 cells) at lower concentrations. Also, silibinin+5-FLU inhibit the cancer stem cell population and significantly reduce the bulk tumor cells [48]. SIL can play a protective role in 5-FLU induced oxidative stress in liver and kidney tissues by eliminates 5-FLU toxicity. The study concluded that this kind of effect is due to the powerful antioxidant and anti-inflammatory properties of SIL [49].

4. Silymarin and Metallic Nanoparticles (gold nanoparticles-GNP, silver nanoparticles-SNP)

The design of composites based on gold nanoparticles (GNPs) and silver nanoparticles (SNPs) functionalized or conjugated with silymarin opens new areas of research in medicine. These metallic nanoparticles are among the most studied in the literature due to their properties, biological activities, photocatalytic hydrogen production and photocatalytic activity toward organic compounds [50, 51]. Also have ability to bind various biomolecules such as amino acids, proteins, anesthetics, antibiotics [52-73]. In this way, they managed to acquire a variety of biomedical applications, including the treatment of cancer and infections that occurred after postoperative surgery in orthopedic and dental surgery. Among the metallic nanoparticles, we remember also platinum nanoparticles that are also useful in biomedical applications but this rare metal are too expensive, so he received less attention [74, 75].

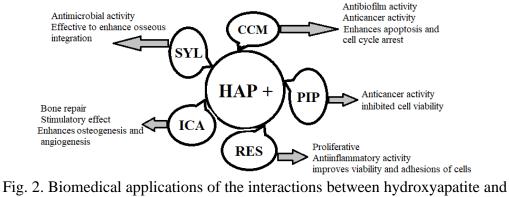
S. Clichici and collaborators loaded the *gold nanoparticles with silymarin*. They observed an improved liver function and reduced cholestasis. The administration of silymarin loaded gold nanoparticles in liver function, significantly decreased the aspartate aminotransferase level (ASAT) in the serum. They concluded that the effect of gold nanoparticles coated with silymarin was significantly better than silymarin alone [76]. On the other hand, S. Staroverov et al examined the silymarin conjugated colloidal gold nanoparticles to see the liver protecting activity. The results show that conjugate administration interfered with glutathione depletion in hepatocytes and stimulated monocyte macrophage function. So, they conclude that SIL-GNP conjugate is possible to be used as a potential liver-protecting drug [77]. A. S. Abdullah and coworkers prepared

Silymarin conjugated gold nanoparticles to improve SIL bioavailability and release for potentiating its antifibrotic action in a rat model intoxicated with CCl₄. They concluded that silymarin conjugated gold nanoparticles may induce antifibrosis effects by enhancing the hepatic expression of the protective microRNAs [78]. In the case of SNP, R. Mohammadinejad synthesized silver nanoparticles using S. marianum extract that can lead to mediate colloidal spherical nanoparticles ranging in size from 1 to 25 nm. Silymarin extracted from seed and fruit is showed to be a good source for synthesis of stable SNPs with simple process of synthesis, low cost and eco-friendly [79, 80].

5. Silymarin and bioactive compounds interaction with hydroxyapatite

Hydroxyapatite is a calcium phosphate found in the hard tissues of the human body such as bones and teeth. In orthopedic and dental surgery is applied for bone grafts or as a covering material for medical implants [81-84], in remineralization of enamel [85, 86] or to remove the heavy metals from wastewater [87]. In terms of implant coverage, it improves cell viability and adhesion, provides good rigidity and low elasticity. To mimic as accurately as possible, the chemical composition of the bone tissue, calcium from hydroxyapatite can be partially replaced with elements such as magnesium, silicon, zinc, strontium in small quantities. The resulting effects led to thermal stability, good solubility, and a favorable response in terms of bone regeneration [88-101].

The interaction between proactive compounds extracted from plants and hydroxyapatite is a new concept and is highlighted in figure 2. The literature studies show that medicine is progressing from one year to the next to find personalized treatments for everyone with medical conditions. In this context, it is desired that patients be treated targeted and less traumatizing as possible so that the beneficial effects are as fast as possible. Although the concept requires extensive daily study for continuous progress in the development of composite materials, the mechanism of action will be elucidated and ready for application.



bioactive compounds extracted from plants

54

Regarding the improvement of *hydroxyapatite with silymarin*, the specialized literature shows that together they have antimicrobial activity against pathogens e.g. Pseudomonas sp, Staphylococcus aureus, Streptococcus mutans, Enterococcus faecalis and Candida albicans [102]. Silymarin is known to promote osteoblast proliferation, inhibits osteoclast proliferation and helps to bone regeneration. So, Z.-S. Tao and his group incorporated silymarin into hydroxyapatite and used it as a coating for titanium implants. They highlight that SIL promotes bone formation around the implant in osteoporotic rats; increases implant osseointegration and improves trabecular microarchitecture. The conclusion of this study is that SIL can be an effective approach to enhance osseous integration of the HAP coated implant in bone [103]. To improve the bioavailability of Silymarin, formulations were developed for oral administration, such as nanoemulsions, nanostructured lipid carrier, solid nanodispersions, inclusion complexation, polymeric and inorganic nanoparticles, liposomes, and solid lipid nanoparticles [104]. An idea that can open new research area is forsterite as an alternative carrier for silvmarin which may allow controlled drug release at the target site [105-114]

Curcumin functionalized hydroxyapatite has anti-biofilm activity tested on human osteoblast-femural cell line (HO-f). W.-H. Lee et al highlight that after 3 days of incubation osteoblast proliferation was lowered. They also show that if incubation period is prolonged to two weeks no significant differences were observed. However, CCM functionalized HAP inhibits bacterial cell attachment and subsequent biofilm maturation stages in both S. aureus and P. Aeruginosa [115]. Another study conducted by Ş. M. Eskitoros-Togay tested CCM-HAP against breast cancer MCF7 cells line. They loaded CCM-HAP into poly (Ecaprolactone)/poly (ethylene oxide) and prepared 7 samples noted PCL/PEO, PCL/PEO/0.1wt.%HAP, PCL/PEO/0.3 wt. %HAP, PCL/PEO/0.5 wt. %HAP, PCL/PEO/0.1 wt. %HAP-CCM, PCL/PEO/0.3 wt. %HAP-CCM, PCL/PEO/0.5 wt. %HAP-CCM. The obtained results of viability of cells show after 24 h of incubation and 94 %, 92 %, and 93 % for PCL/PEO/0.1HAP, PCL/PEO/0.3HAP, and PCL/PEO/0.5HAP respectively. After 48 h incubation 93 %, 89 % and 90 %. In the case of PCL/PEO/0.1 wt. %HAP-CCM, PCL/PEO/0.3 wt. %HAP-CCM, PCL/PEO/0.5 wt. %HAP-CCM [116].

W.-H. Lee et al also study this kind of composite, CCM-HAP but the surface of HAP was functionalized with different carboxylic acids to harbor negative charges and increase drug (CCM-NPs) loading capacity of HAP. Their conclusion was that CCM loaded carboxylic acids-HAPs on MCF-7 cells were directly correlated to the release rate of CCM nanoparticles from HAP carrier, depending on the types of carboxylic acids used. This type of composite showed higher anti-

cancer activity and resulted in enhanced apoptosis and cell cycle arrest compared to unmodified HAP [117].

K. AbouAitah and his group prepared nanoformulations consisting in aggregates of *HAPs loaded with PIP* and tested in vitro against HCT116 cells (colon cancer), MCF7(human breast adenocarcinoma) and Caco2 (human colon carcinoma cells) and WI-38 (human fibroblasts cells. The obtained results show potential for targeting this kind of colon cancer cells, but a strong reduction was also shown on MCF7 cells when treated with HAP-PIP at pH 9.3, 200 μ l incubated for 72h. Unloaded nanoparticles exhibited weak cytotoxicity towards WI-38 fibroblasts. They concluded that the cell viability of cancers depended on cell line, concentration, incubation time, and delivery method of PIP. Also, increasing the incubation time from 48 h to 72 h inhibited the cell viability of all investigated cells [118].

Resveratrol (RES) and hydroxyapatite (HAP) composite is known to have proliferative activity in the case of human adipose-derived mesenchymal stromal stem cells (hASCs). nHAP and RES improve the adhesion and spreading of cells, improve viability, metabolic activity, and mitochondrial potential. The composite has a great pharmacological potential as carrier for bioactive compound delivery [119]. In the case of RAW264.7 cells (leukemia cells in mouse macrophage cell line) the researchers synthesized nano-hydroxyapatite (n-HA)/resveratrol (Res)/chitosan (CS) microspheres. They obtained antiinflammatory activity evidenced by the decreased expression of pro-inflammatory cytokines TNF- α , IL-1 β and iNOS in RAW264.7 cells in a dose dependent manner. This kind of composite could stimulate BMSCs proliferation and osteodifferentiation, as well bone remodeling under osteoporotic condition [120].

Icariin loaded on micro/nano HAP granules was tested on rat femoral defect model on bone mesenchymal stem cells (BMSCs) at two concentrations 200 μ M and 2000 μ M. The conclusion of study was that icariin could promote the osteogenic differentiation and expression of angiogenic factors of BMSCs. At concentration of 200 μ M, icariin had the strongest stimulatory effect but in 2000 μ M could enhance both osteogenesis and angiogenesis in vivo. They evidence that a small molecule of icariin could penetrate HAP network that could slow down the release rate in the 2000 μ M group [121]. Icariin was also loaded in chitosan/nano-sized hydroxyapatite (IC–CS/HA) by J. Fan et al. They study the bone repairing, more specify the bone marrow derived stroma cells (BMSCs). The loaded icariin in this composition demonstrates the stimulation of bone marrow derived stroma cell alkaline phosphatase activity and formation of mineralized nodules [122].

Regarding *Carotenoids and hydroxyapatite*, the literature presents the potential use of hydroxyapatite nanoparticles as adsorbent for β -carotene/lycopene. S. Kongsri et al demonstrate through adsorption isotherms that

56

the adsorption behaviours of the carotenoids on hydroxyapatite nanoparticles and chitosan substrates were well fitted [123]. They found that the adsorption capacity values for lycopene are greater than those of β -carotene for both hydroxyapatite nanoparticles and chitosan. They say that hydroxyapatuite can be a good substrate for carotenoids adsorption in physiological functions. In other study conducted also from S. Kongsri et al investigate the adsorption of carotenoid from tomato extract using nanocrystalline fish hydroxyapatite in the presence of sodium dodecyl sulfate. Adsorption interactions are possible through various mechanisms such as ion exchange, hydrophobic interaction and electrostatic forces. In this sense, the carotenoid can be adsorbed on fish hydroxyapatite surface through hydrophobic interactions [124].

This is how the incorporation of plant compounds into hydroxyapatitebased composites can bring about improvements in the treatment of various types of cancer cells or even in the case of bone regeneration. It is a broad idea that requires continuous development and intense application on the cells to obtain results with desired effects.

6. Silymarin and bioactive compounds interaction with carotenoid

Carotenoids are natural pigments that can protect the body against various diseases, enhance the immune system, and play a role in cell membrane stability, photosynthesis, and cellular differentiation. They also contribute to health benefits through anti-cancer, anti-inflammatory, anti-bacterial, anti-diabetic and neuroprotective activity [125-128]. Carotenoids are found in plants such as vegetables, fungi, algae, or bacterial species and can also prevent atherosclerosis, age-related macular degeneration, and other chronic diseases [129-131]. Two groups of carotenoids exist: xanthophylls (Canthaxanthin, Zeaxanthin, Lutein, Astaxanthin) [132-137] and carotenes (β -carotene, Lycopene) [138, 139] both having antioxidant properties.

Specialized literature shows that carotenoids films can be studied by Langmuir-Blodgett Technique. Some studies conducted by M. Tomoaia-Cotisel and coworkers produced self-assembled and supra-molecular structures at air/water, oil/water, benzene/water and gas/liquid interfaces [140-155]. This approach opened new potentials of research in monolayers and multilayers structures with biological and biomedical importance for life science. The interaction with various biomolecules such that mentioned in this review, may be explored by this technique that can simulate the cell membrane using a single layer oriented at air/water interfaces. The interest to develop new biomaterials for regenerative medicine, tissue engineering, cancer therapy, self-assembled scaffolds and drug delivery make from Langmuir Blodgett Technique (LBT) an essential instrumentation in manufacture of thin film in which the surface potential of biomolecules can be measured to characterize the miscibility. The technique is useful to biomedical applications by nanoscale interactions. Such applications are based on self-assemblies of collagen [156-158], lipids, phospholipids and galactolipids [159-165], antioxidants [166-169], fatty acids [170-173], proteins [174, 175], cholesterol and lecithin [176-178], drugs [179-181].

The literature highlights study the orientation and localization of carotenoids in the lipid membrane. A schematic representation of carotenoids that are hydrophobic molecules can be seen in Figure 3.

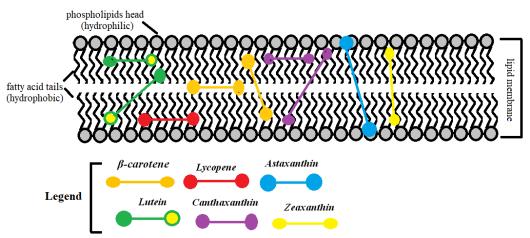


Fig. 3. Localization and orientation of carotenoids in lipid membrane

Carotenoids can incorporate into lipid membrane starting from the strongest to the weakest. C. Tan et al shows that liposomes can act as a delivery system for various carotenoids, so they can display different loading ability into lipid bilayer [182]. The macular carotenoid is oriented perpendicularly to the membrane surface which ensures high solubility and stability [183]. J. Widomska and collaborators highlighted that macular xanthophyll interact with proteins and lipid from membrane to absorb light energy, modulate oxidative stress and influence signal transduction cascades [184].

Lycopene ($C_{40}H_{56}$) is a carotenoid found abundant in tomato but also in small amounts in guava, pink grapefruit, papaya, and watermelon. Is a lipophilic red color pigment that derived from an acyclic structure with 13 carbon double bonds (11 conjugated double bonds and 2 non-conjugated double bonds) arranged in a linear array. It is insoluble in water and soluble in organic solvents. As can be seen in figure 3, lycopene is oriented parallel to cell membrane surface within the lipid bilayer, and it is expected to be poor hydrophilic antioxidant due to its limited interaction with aqueous phase radicals in the lipid bilayer [185].

The literature shown that lycopene can arouse the proliferation of osteoblast-like SaOS-2 cells (a human osteosarcoma cell line which displays

several osteoblastic features) and has inhibitory effect on MC3T3 cells proliferation; MC3T3 cell line is an osteoblast precursor cell line derived from mouse calvaria. Also, lycopene may be beneficial in cardiovascular diseases and ulcers; as an antioxidant, it can prevent the oxidative damage of DNA, lipids and proteins and induces apoptotic cell death. It seems that lycopene also decreased Bcl-2 and increased levels of Bax, inhibited phosphorylation of extracellular signal-regulated kinase, and it may protect against the development of gastric cancer [186-188].

The β -carotene is the most abundant carotenoid in the human body and derived from the acyclic structure, C₄₀H₅₆, possesses a long chain of conjugated double bonds and two β -iononic rings. The β -carotene and lycopene orientation in the lipid membrane, as seen in figure 3, can be governed by van der Waals interactions with the hydrocarbon acyl chains of lipid molecules, forming the hydrophobic core of the membrane [189]. Is found in most vegetables and fruits such as carrots, spinach, kale, parsley, summer squash, tomato's, sweet potatoes, broccoli, and mango. It has the highest bioactivity and acts as a precursor to produce vitamin A. β-Carotene can be partially converted to vitamin A, but the unconverted β -carotene is incorporated in chylomicrons, secreted into the lymph, and then transported to the liver. β -Carotene can reduce the risk of osteoporosis. As a mechanism in cancer prevention β -carotene acts as a pro-oxidant in leukemia cells (HL-60) and colon adenocarcinoma cells (LS-174 and WiDr). Also, in lung cancer/lung carcinoma cell (A549) increase the oxidative stress marker (8-oxodG). Inhibit cancer cell growth by increased production of ROS via activation of NF-kB in leukemia and colon cancer/human leukemic cells (HL-60), colon adenocarcinoma cells (LS-174 and WiDr) and activate apoptosis in Leukemia/HL-60 cells. In gastric cancer/AGS cells β -carotene increased levels of caspase 3, ROS, cytochrome c and Bax (proapoptotic effector molecules from Bcl-2 family) [190-193].

Lutein is a 40-carbon hydroxylated carotenoid with β and ε type ionone ring and acts as a pro-oxidant in breast cancer and increases the levels of phosphorylated p53 and heat shock protein 60 [190, 193-195]. Its orientation in cell membrane can be seen in figure 3. X. Gong et al, in their study highlighted that the treatment with lutein on MDA-MB-468 and MCF-7 cells inhibited cell cycle progression. An increased population of cells in G1 phase, a reduction in G2 phase in MDA-MB-468 cells, as well as a decreased cell population in G1 phase and an increase in G2 phase in MCF-7 cells was observed [196]. It can be found abundant in marigold flowers, broccoli, lettuce, cilantro, kale, and sweet potato but also in small amount in pepper, maize, black palm, and pumpkin [197].

Astaxanthin with molecular formula $C_{40}H_{52}O_4$ consists of two terminal rings joined by a polyene chain [198]. It has two asymmetric carbons located at the 3, 3' positions of the β -ionone ring with hydroxyl group (-OH) on either end of

the molecule. The position in cell membrane can be seen in figure 3, the terminal ring is able to scavenge radicals at the surface and in the interior of phospholipids membrane. Its membrane insertion is due to its linear molecular appearance and extends across the entire width of the lipid membrane [199, 200]. It is a red pigment found naturally in shrimp, crab, and salmon, is insoluble in water but soluble in most organic solvents. It is a keto-carotenoid synthesized by plants and microorganisms [201]. In the case of cancer treatment, astaxanthin significantly inhibited prostate cancer DU145 cells proliferation and promotes the apoptosis of these cells.

In the literature is highlighted that astaxanthin effectively inhibits the cloning ability of DU145 cells, downregulates the gene expression of JAK2, BCL-2, NF-kB and upregulates the gene expression of BAX, Caspase3 and Caspase9 [202]. In the case of human glioblastoma cell line U251MG, astaxanthin suppressed cell viability at concentrations of more than 1 and 0.1 μ M. It is shown that the treatment for 48 h decreased the expression of cyclinD1 and increased the expression of p27 [203].

Zeaxanthin is a yellow-orange xanthophyll also known as β , β -carotene-3,3'-diol which contain two hydroxyl groups with a higher polarity. The presence of the two hydroxyl groups determines its orientation in lipid membrane, figure 3, and enhances the stability of zeaxanthin [183]. It has been reported by literature to exhibit cytotoxic effects. It is a lipophilic compound insoluble in aqueous media. It can induce G2/M cell cycle and apoptosis in gastric cancer cells by upregulating Bax pro-apoptotic factor and down-regulating Bcl-2 anti-apoptotic proteins. Zeaxanthin has been shown to have protective effects in eyes (prevents AMD and cataract), liver (reduce ROS and protect against nonalcoholic fatty liver disease), skin (inhibits sunburn) and arteries (protects against atherosclerosis) [204].

It was demonstrated by in vivo and in vitro results that zeaxanthin and lutein has protective effects against chronic eye and cardiovascular diseases, such as age-related macular degeneration (AMD), cataract, coronary heart disease, and stroke [205].

Canthaxanthin (β , β -carotene-4,4'-dione) with (C₄₀H₅₂O₂) is a red-orange xanthophyll's that has nine conjugated double-bonds terminated by two oxo substituents at positions 4 and 4' of the β -ionone backbone. Its orientation in lipid membrane, figure 3, is roughly perpendicularly to the surface of the membrane but also can be oriented parallel to the membrane, localized in the headgroup region. Canthaxanthin can modify the properties of membrane such as promote extended conformation of alkyl lipid chains, can modify the surface of lipid membrane in gel state and promote the aggregation of lipid vesicles [206]. It is used in cosmetics as a natural tanning agent who produces an orange-brown colour to skin. In vivo results highlighted that canthaxanthin led to decrease in

lipid peroxidation by preventing liver DNA damage, enhancing the antioxidant defense in rat liver and increased the activity of alkaline phosphatase [207]. A study conducted by P. Palozza and coworkers, it is indicating that canthaxanthin was able to inhibit the growth of malignant human cell lines, such as WiDr colon adenocarcinoma and SK-MEL-2 melanoma. Also, it induces apoptosis of these cells at the highest dose at the longest time of exposure (48h) [208].

Silymarin and lycopene investigation was conducted by L Garavaglia and coworkers where this combination was administrating in periparturient dairy cows. This combination could be a feed supplement with hepatoprotective and antioxidant activity. The treated cows show a higher milk yield at the beginning of lactation than untreated animals. The synergy effect can contribute to mitigate the negative effects on metabolic adaptation to the lactation [209]. In vitro study demonstrates the protective effect of *beta-carotene and silymarin* on DNA damage induced by L-arginine in lymphocyte culture [210].

R. P. Assis et al, in their study revealed that carotenoids maintain the benefit provided by curcumin alone. The co-administration produces effects on the increase in superoxide dismutase activity. *Curcumin and lycopene* in the case of diabetic rats is more effective in reducing glycemia than carotenoid treatments alone; it also led to significant body weight gain and provides benefits against lipoperoxidation [211]. *Curcumin and lutein* are beneficial in treatment of fecal oocyst in chickens. Their combination enhanced cellular and humoral immunity [212].

B. M. Steiner et al, in their research tried to resolve the lutein strong hydrophobicity and poor chemical stability when it is introduced in many foods. In this sense, some nanoemulsions was prepared that contain Casein-dextran Maillard conjugates, *resveratrol* and GSO (grape seed oil). Casein-dextran improves the physical resistance of nanoemulsions while resveratrol and GSO decreased *lutein* degradation and are effective at improving their chemical stability [213].

A. Kawamura and his group study the effect of anabolic nutrient-rich foods on muscle adaptation induced by resistance training. So, they highlighted that the combined intake of *astaxanthin*, β -carotene, and resveratrol can accelerate protein anabolism in the skeletal muscle of mice. Also, they concluded that even in a small amount this kind of combination can promote protein synthesis during the muscle hypertrophic process after atrophy [214, 215].

In the case of piperine and icariin interactions with carotenoids, information is limited or none. This is an approach that needs special attention to understand the mechanism of action. Beneficial results from this type of combination will exist from the moment when experts in this field will publish their obtained experimental data. Biodegradable material such as calcium sulfate hemihydrate and composites based on gypsum have been used as local drug delivery systems or as construction materials in the living and non-living world [216].

All the data from this section require more details but the literature is limited once again. It seems that interactions of phytochemicals need someone to research this area to find information with medical potential.

7. Conclusions

This review is focused on Silymarin interaction with phytochemicals extracted from plants and their interactions with each other and with hydroxyapatite and carotenoids. Their efficiency was demonstrated in various cancer cells lines such as MCF-7 (breast cancer), bladder carcinoma, hepatocellular carcinoma, colon cancer (HCT116) and against pathogens (S. aureus and P. Aeruginosa). Also is debated the Silymarin combination with chemotherapeutic drugs (doxorubicin, paclitaxel, and 5-fluorouracil) and metallic nanoparticles such as gold and silver.

Herbal therapy demonstrates multiple biological activities in the treatment of chronic diseases and cancer (the cruelest disease in the world that causes millions of deaths a year regardless of age). The presence of medicinal plants in the world has been studied since ancient times and through the mechanism of action that has been intensively studied lately, it has been possible to prevent and maintain the balance between health and death.

However, this approach requires the design and development of new strategies and innovative nanomaterials not only in the treatment of cancer but also in bone regeneration, tissue engineering and viruses that are constantly beginning to appear.

Acknowledgment

This work was supported by grants of the Ministry of Research, Innovation and Digitization, *CNCS/CCCDI-UEFISCDI*, project number 186 and 481, within *PNCDI III*.

Notations and/or Abbreviations

CCM – curcumin, SIL –Silymarin, PIP – piperine, RES – Resveratrol, ICA – Icariin, HAP – hydroxyapatite, GNPs – gold nanoparticles, SNPs – Silver nanoparticles, DOX – doxorubicin, PCT – paclitaxel, 5-FLU – 5-fluorouracil, LBT – Langmuir Blodgett Technique

REFERENCES

[1] G.-A. Paltinean, S. Riga, Gh. Tomoaia, A. Mocanu, M. Tomoaia-Cotisel, Bioactive Compounds from Plants Used as Therapeutic Agents in Biomedical Applications - A Literature Review, Academy of Romanian Scientists, Annals Series on Biological Sciences, **10**(2), 103-141 (2021).

[2] Y. J. Surh, S. S. Han, Y. S. Keum, H. J. Seo, S. S. Lee, Inhibitory effects of curcumin and capsaicin on phorbol ester-induced activation of eukaryotic transcription factors, NF-kappaB and AP-1, Biofactors, **12**(1–4), 107–112 (2000).

[3] T. Dorai, Y. C. Cao, B. Dorai, R. Buttyan, A. E. Katz, Therapeutic potential of curcumin in human prostate cancer. III. Curcumin inhibits proliferation, induces apoptosis, and inhibits angiogenesis of LNCaP prostate cancer cells in vivo, Prostate, **47**(4), 293–303 (2001).

[4] S. Garg, A. Garg, Encapsulation of curcumin in silver nanoparticle for enhancement of anticancer drug delivery, Int. J. Pharm. Sci. Res., **9**(3), 1160-1166 (2018).

[5] D. Perrone, F. Ardito, G. Giannatempo, M. Dioguardi, G. Troiano, L. Lo Russo, A. De Lillo, L. Laino, L. Lo Muzio, Biological and therapeutic activities, and anticancer properties of curcumin (Review), Exp. Ther. Med., **10**, 1615-1623 (2015).

[6] L. Rácz, M. Tomoaia-Cotișel, Cs.-P. Rácz, P. Bulieris, I. Grosu, S. Porav, A. Ciorîță, X. Filip, F. Martin, G. Serban, I. Kacso, Curcumin-Whey protein solid dispersion system with improved solubility and cancer cell inhibitory effect, Stud. Univ. Babes-Bolyai Chem, **66**(3), 209-224 (2021).

[7] L. Z. Racz, Cs. P. Racz, L.-C. Pop, Gh. Tomoaia, A. Mocanu, I. Barbu, M. Sárközi, I. Roman, A. Avram, M. Tomoaia-Cotisel, V.-A. Toma, Strategies for Improving Bioavailability, Bioactivity, and Physical-Chemical Behavior of Curcumin, Molecules, 27(20), 6854 (2022).

[8] L. Z. Racz, Cs. P. Racz, O. Horovitz, Gh. Tomoaia, A. Mocanu, I. Kacso, M. Sarkozi, M. Dan, S. Porav, G. Borodi, M. Tomoaia-Cotisel, Complexation of curcumin using whey proteins to enhance aqueous solubility, stability and antioxidant property, Stud. Univ. Babes-Bolyai Chem., **67**(3), 75-99 (2022).

[9] L. Z. Racz, G.-A. Paltinean, I. Petean, Gh. Tomoaia, L.- C. Pop, G. Arghir, E. Levei, A. Mocanu, Cs.-P. Racz, M. Tomoaia-Cotisel, Curcumin and Whey Protein Binding And Structural Characteristics Of Their Complex Evidenced By Atomic Force Microscopy, Stud. Univ. Babes-Bolyai Chem., **67**(3), 61-74 (2022).

[10] A. Tajmohammadi, B. M. Razavi, H. Hosseinzadeh, Silybum marianum (milk thistle) and its main constituent, silymarin, as a potential therapeutic plant in metabolic syndrome: A review, Phytother. Res., **32**(10), 1933-1949 (2018). <u>https://doi.org/10.1002/ptr.6153</u>.

[11] Neha, A. S. Jaggi, N. Singh, *Silymarin and its role in chronic diseases*, in book Drug Discovery from Mother Nature, Advances in Experimental Medicine and Biology 929, edited by S. C. Gupta, S. Prasad, B. B. Aggarwal (Springer International Publishing, Switzerland, 2016), pp. 25-44.

[12] G. A. Paltinean, Gh. Tomoaia, S. Riga, A. Mocanu, M. Tomoaia-Cotisel, Sylimarin based complexes – a mini review, Annals Series on Biological Sciences, **11**(1), 146-166 (2022).

[13] M. Saeed, D. Babazadeh, M. Arif, M. A. Arain, Z. A. Bhutto, A. H. Shar, M. U. Kakar, R. Manzoor, S. Chao, Silymarin: a potent hepatoprotective agent in poultry industry, World's Poult. Sci. J., 73, 483-492 (2017).

[14] S. K. Das, S. Mukherjee, D. M. Vasudevan, Medicinal properties of milk thistle with special reference to Silymarin – An owerview, Nat. Prod. Radiance, **7**(2), 182-192 (2008).

[15] Y. Haddad, D. Vallerand, A. Brault, P. S. Haddad, Antioxidant and hepatoprotective effects of silibinin in a rat model of nonalcoholic steatohepatitis. Evid. Based Complement Altern. Med., 2011, Article ID 647903 (2011). <u>https://doi.org/10.1093/ecam/nep164</u>

[16] C. P. Colturato, R. P. Constantin, A, S. Maeda Jr, R. Polimeni, C. Nair, S. Yamamoto, A. Bracht, E. L. Ishii-Iwamoto, J. Constantin, Metabolic effects of silibinin in the rat liver, Chem. Biol. Interact., 195(2), 119–132 (2012).

[17] S. Bhattacharya, *Milk thistle (Silybum marianum L. Gaert.) seeds in health*, In book: Nuts and seeds in health and disease prevention, eds. V. R. Preedy, R. R. Watson, V. Patel, (Academic Press, London, UK, 2011), chapter 90, pp. 759-766.

[18] R. Gazak, D. Walterova, V. Kren, Silybin and silymarin—new and emerging applications in medicine, Curr. Med. Chem., **14**, 315–338 (2007).

[19] D.-Y. Ouyang, L.-H. Zeng, H. Pan, L.-H. Xu, Y. Wang, K.-P. Liu, X.-H. He, Piperine inhibits the proliferation of human prostate cancer cells via induction of cell cycle arrest and autophagy, Food Chem Toxicol., **60**, 424-430 (2013).

[20] J. Zhang, X. Zhu, H. Li, B. Li, L. Sun, T. Xie, T. Zhu, H. Zhou, Z. Ye, Piperine inhibits proliferation of human osteosarcoma cells via G2/M phase arrest and metastasis by suppressing MMP-2/-9 expression, Int. Immunopharmacol., **24**(1), 50-58 (2015).

[21] S. Banerjee, P. Katiyar, V. Kumar, S. S. Saini, R. Varshney, V. Krishnan, D. Sircar, P. Roy, Black pepper and piperine induce anticancer effects on leukemia cell line, Toxicol. Res., 10(2), 169–182 (2021).

[22] L. Guo, Y. Yang, Y. Sheng, J. Wang, S. Ruan, C. Han, Mechanism of piperine in affecting apoptosis and proliferation of gastric cancer cells via ROS-mitochondria-associated signalling pathway, J. Cell Mol. Med., **25**(20), 9513-9522 (2021).

[23] Y. Bai, Q.-Q. Mao, J. Qin, X.-Y. Zheng, Y.-B. Wang, K. Yang, H.-F. Shen, L.-P. Xie, Resveratrol induces apoptosis and cell cycle arrest of human T24 bladder cancer cells in vitro and inhibits tumor growth in vivo, Cancer Sci., **101**(2), 488-493 (2010).

[24] M. Yousef, I. A. Vlachogiannis, E. Tsiani, Effects of resveratrol against lung cancer: In Vitro and in vivo studies, Nutrients, 9(11), 1231 (2017). doi: 10.3390/nu9111231.

[25] A. A. Sprouse, B.-S. Herbert, Resveratrol augments Paclitaxel treatment in MDA-MB-231 and Paclitaxel-resistant MDA-MB-231 breast cancer cells, Anticancer Res., 34(10), 5363-5374 (2014).

[26] Q. Xu, L. Zong, X. Chen, Z. Jiang, L. Nan, J. Li, W. Duan, J. Lei, L. Zhang, J. Ma, X. Li, Z. Wang, Z. Wu, Q. Ma, Z. Ma, Resveratrol in the treatment of pancreatic cancer, Ann. N. Y. Acad. Sci., **1348**(1), 10-19 (2015).

[27] X. Wu, W. Kong, X. Qi, S. Wang, Y. Chen, Z. Zhao, W. Wang, X. Lin, J. Lai, Z. Yu, G. Lai, Icariin induces apoptosis of human lung adenocarcinoma cells by activating the mitochondrial apoptotic pathway, Life Sci., **239**, 116879 (2019). DOI: 10.1016/j.lfs.2019.116879.

[28] Z.-F. Gu, Z.-T. Zhang, J.-Y. Wang, B.-B. Xu, Icariin exerts inhibitory effects on the growth and metastasis of KYSE70 human esophageal carcinoma cells via PI3K/AKT and STAT3 pathways, Environ. Toxicol. Pharmacol., **54**, 7-13 (2017).

[29] C. Zhang, X. Sui, Y. Jiang, X. Wang, S. Wang, Antitumor effects of icaritin and the molecular mechanisms, Discov. Med., **29**(156), 5-16 (2020).

[30] Who Report On Cancer: Setting priorities, investing wisely and providing care for all. Geneva: World Health Organization; 2020. Licence: CC BY-NC-SA 3.0 IGO.

[31] CureAll framework: WHO Global Initiative for Childhood Cancer. Increasing access, advancing quality, saving lives. Geneva: World Health Organization; 2021. Licence: CC BY-NC-SA 3.0 IGO; https://creativecommons.org/licenses/by-nc-sa/3.0/igo/

[32] A. Montgomery, T. Adeyeni, K. K. San, R. M. Heuertz, U. R. Ezekiel, Curcumin sensitizes silymarin to exert synergistic anticancer activity in colon cancer cells, J. Cancer, **7**(10), 1250-1257 (2016).

[33] N. Abdel-Magieda, A. A. Elkady, Possible curative role of curcumin and silymarin against nephrotoxicity induced by gamma-rays in rats, Exp. Mol. Pathol., **111**, 104299 (2019). https://doi.org/10.1016/j.yexmp.2019.104299

[34] M. M. Ahmad, N. A. Rezk, A. Fawzy, M. Sabry, Protective effects of curcumin and silymarin against paracetamol induced hepatotoxicity in adult male albino rats, Gene, **712**, 143966 (2019). https://doi.org/10.1016/j.gene.2019.143966

[35] M. Nasiri, N. Zarghami, K. Nejati Koshki, M. Mollazadeh, M. P. Moghaddam, M. R. Yamchi, R. J. Esfahlan, A. Barkhordari, A. Alibakhshi, Curcumin and Silibinin Inhibit Telomerase Expression in T47D Human Breast Cancer Cells, Asian Pac. J. Cancer Prev., **14**(6), 3449-3453 (2013).

[36] A. Farzanegan, M. Shokuhian, S. Jafari, F. S. Shirazi, M. Shahidi, Anti-histaminic Effects of Resveratrol and Silymarin on Human Gingival Fibroblasts, Inflammation, 42(5), 1622-1629 (2019).

[37] M. Shahidi, F. Vaziri, A. Haerian, A. Farzanegan, S. Jafari, R. Sharifi, F. S. Shirazi, Proliferative and Anti-Inflammatory Effects of Resveratrol and Silymarin on Human Gingival Fibroblasts: A View to the Future, J. Dent. (Tehran), **14**(4), 203-211 (2017).

[38] W.-C. Hsieh, C.-W. Yang, Y.-S. Haung, T.-W. Chao, T.-F. Tsai, I.-J. Su, Chemoprevention of HBV-related hepatocellular carcinoma by the combined product of resveratrol and silymarin in transgenic mice, Func. Foods Health Dis., **3**(9), 341-352 (2013).

[39] R. Shukla, S. J. Surana, A. U. Tatiya, S. K. Das, Investigation of hepatoprotective effects of piperine and silymarin on Dgalactosamine induced hepatotoxicity in rats, Res. J. Pharm., Biol. Chem. Sci., **2**(3), 975-982 (2011).

[40] S. Javed, W. Ahsan, K. Kohli, Pharmacological influences of natural products as bioenhancers of silymarin against carbon tetrachloride-induced hepatotoxicity in rats, Clin. Phytosci., 4(1), 18 (2018) <u>https://doi.org/10.1186/s40816-018-0079-6</u>.

[41] E. Yurtcu, O. D. Iseri, F. I. Sahin, Genotoxic and cytotoxic effects of doxorubicin and silymarin on human hepatocellular carcinoma cells, Hum. Exp. Toxicol., **33**(12), 1269-1276 (2014).

[42] N. Patel, C. Joseph, G. B. Corcoran, S. D. Ray, Silymarin modulates doxorubicin-induced oxidative stress, Bcl-xL and p53 expression while preventing apoptotic and necrotic cell death in the liver, Toxicol. Appl. Pharmacol., **245**(2), 143-152 (2010).

[43] E. Cecen, T. Dost, N. Culhaci, A. Karul, B. Ergur, M. Birincioglu, Protective effects of silymarin against doxorubicin-induced toxicity, Asian Pac. J. Cancer Prev., **12**(10), 2697-2704 (2011).

[44] F. Gheybi, S. H. Alavizadeh, S. M. Rezayat, E. Zendedel, M. Jaafari, Chemotherapeutic activity of Silymarin combined with doxorubicin liposomes in 4T1 breast cancer cells, Nanomed. Res. J., **4**(1), 29-34 (2019).

[45] A. K. Tyagi, R. P. Singh, C. Agarwal, D. C. F. Chan, R. Agarwal, Silibinin strongly synergizes human prostate carcinoma DU145 cells to doxorubicin-induced growth Inhibition, G2-M arrest, and apoptosis, Clin. Cancer Res., **8**(11), 3512-3519 (2002).

[46] Y. Zhang, Y. Ge, X. Ping, M. Yu, D. Lou, W. Shi, Synergistic apoptotic effects of silibinin in enhancing paclitaxel toxicity in human gastric cancer cell lines, Mol. Med. Rep., **18**(2), 1835-1841 (2018).

[47] O. Molavi, F. Narimani, F. Asiaee, S. Sharifi, V. Tarhriz, A. Shayanfar, M. Hejazi, R. Lai, Silibinin sensitizes chemo-resistant breast cancer cells to chemotherapy, Pharm. Biol.**55**(1),729-739 (2017).

[48] S. Patel, B. Waghela, K. Shah, F. Vaidya, S. Mirza, S. Patel, C. Pathak, R. Rawal, Silibinin, A Natural Blend In Polytherapy Formulation For Targeting Cd44v6 Expressing Colon Cancer Stem Cells, Sci. Rep., 8, 16985 (2018). DOI:10.1038/s41598-018-35069-0

[49] E. Sengul, V. Gelen, S. Yildirim, E. Senturk, Y. Dag, G. Eser, M. Gok, Investigation of Effects of Silymarin in 5- Fluorouracil Hepatotoxicity and Nephrotoxicity in Mice, (2021) DOI: https://doi.org/10.21203/rs.3.rs-448267/v1.

[50] Z. Pap, E. Karácsonyi, L. Baia, L.-C. Pop, V. Danciu, K. Hernádi, K. Mogyorósi, A. Dombi, TiO2/WO3/Au/MWCNT composite materials for photocatalytic hydrogen production: Advantages and draw-backs, Phys. Status Solidi B, **12**, 2592-2595 (2012).

[51] E. Karácsonyi, L. Baia, A. Dombi, V. Danciu, K. Mogyorósi, L.-C. Pop, G. Kovács, V. Coşoveanu, A. Vulpoi, S. Simon, Zs. Pap, The photocatalytic activity of TiO2/WO3/noble metal (Au or Pt) nanoarchitectures obtained by selective photodeposition, Catal. Today, **208**, 19-27 (2013).

[52] O. Horovitz, A. Mocanu, Gh. Tomoaia, M. Crisan, L. D. Bobos, C. Racz, M. Tomoaia-Cotisel, Amino acids binding to gold nanoparticles, Stud. Univ. Babes-Bolyai Chem., 52(3), 53-71 (2007). [53] A. Mocanu, I. Cernica, Gh. Tomoaia, L. D. Bobos, O. Horovitz, M. Tomoaia-Cotisel, Self-assembly characteristics of gold nanoparticles in the presence of cysteine, Colloid Surf. A, 338(1-3), 93-101 (2009).

[54] I. Petean, Gh. Tomoaia, O. Horovitz, A. Mocanu, M. Tomoaia-Cotisel, Cysteine mediated assembly of gold nanoparticles, J. Optoelectron. Adv. Mat., **10**(9), 2289-2292 (2008).

[55] O. Horovitz, A. Mocanu, Gh. Tomoaia, L. D. Bobos, D. Dubert, I. Daian, T. Yupsanis, M. Tomoaia-Cotisel, Lysine mediated assembly of gold nanoparticles, Stud. Univ. Babes-Bolyai Chem., **52**(1), 97-108 (2007).

[56] O. Horovitz, Gh. Tomoaia, A. Mocanu, T. Yupsanis, M. Tomoaia-Cotisel, Protein binding to gold colloids, Gold Bull., **40**(3), 213-218 (2007).

[57] Gh. Tomoaia, P. T. Frangopol, O. Horovitz, L. D. Bobos, A. Mocanu, M. Tomoaia-Cotisel, The effect of arginine on gold nanoparticles in colloidal solutions and in thin films, J. Nanosci. Nanotechnol., **11**(9), 7762-7770 (2011).

[58] O. Horovitz, Gh. Tomoaia, A. Mocanu, T. Yupsanis, M. Tomoaia-Cotisel, Protein binding to gold auto assembled films, Gold Bull., **40**(4), 295-304 (2007).

[59] A. Mocanu, R. D. Pasca, Gh. Tomoaia, A. Avranas, O. Horovitz, M. Tomoaia-Cotisel, Selective effect of procaine tetracaine and dibucaine on gold nanoparticles, J. Nanosci. Nanotechnol., 12(12), 8935-8939 (2012).

[60] M. Tomoaia-Cotisel, A. Mocanu, O. Horovitz, E. Indrea, Gh. Tomoaia, I. Bratu, *Self-assembly of gold nanoparticles functionalized with amino acids and aleurone globular protein*, in Book series (Proceedings of SPIE): Advanced Topics in Optoelectronics, Microelectronics, and Nanotechnologies IV, edited by P. Schiopu, C. Panait, G. Caruntu, A. Manea, (Constanta, Romania, 2009), **Vol. 7297**, Article No: UNSP 729708. doi: 10.1117/12.823616.

[61] R. D. Pasca, A. Mocanu, S. C. Cobzac, I. Petean, O. Horovitz, M. Tomoaia-Cotisel, Biogenic syntheses of gold nanoparticles using plant extracts, Particul. Sci. Technol., **32**(2), 131-137 (2014).

[62] A. Mocanu, O. Horovitz, Cs. P. Racz, M. Tomoaia-Cotisel, Green synthesis and characterization of gold and silver nanoparticles, Rev. Roum. Chim., **60**(7-8), 721-726 (2015).

[63] O. Horovitz, A. Mocanu, Gh. Tomoaia, L. Olenic, O. Mihailescu, G. Borostean, A. Popoviciu, C. Craciun, T. Yupsanis, M. Tomoaia-Cotisel, *Synthesis, characterization and properties of gold nanoparticles in colloidal aqueous solutions in the absence and in the presence of globular proteins. Auto-assembled gold nanostructures in thin films*, in Convergence of micro-nanobiotechnologies, Series in Micro and Nanoengineering, Eds: M. Zaharescu, E. Burzo, L.

Dumitru, I. Kleps, D. Dascalu (Romanian Academy Press, Bucharest, Romania, 2006), vol. 9, pp. 132-146.

[64] O. Horovitz, A. Mocanu, Gh. Tomoaia, C. Craciun, M. Tomoaia-Cotisel, *Assembly of gold nanoparticles mediated by amino acids*, in Progress in Nanoscience and Nanotechnologies, Series in Micro and Nanoengineering, eds I. Kleps, A. C. Ion, D. Dascalu (Romanian Academy Press, Bucharest, Romania, 2007), vol. 11, pp. 23-36.

[65] L. Barbu-Tudoran, Gh. Tomoaia, O. Horovitz, A. Mocanu, M. Tomoaia-Cotisel, Selfassembly characteristics of gold nanoparticles in the presence of arginine, J. Optoelectron. Adv. M., 10(9), 2293-2297 (2008).

[66] M. Tomoaia-Cotisel, *Multifunctional nanostructures formed of gold or silver nanoparticle and different biomolecules with medical applications*, (e-Book, Cluj University Press, Cluj-Napoca, Romania, 2016) pp. 1-322.

[67] A. Avram, Gh. Tomoaia, A. Mocanu, M. Tomoaia-Cotisel, Gold nanoparticles and chemotherapeutic agents, Annals of the Academy of Romanian Scientists, Series on Physics and Chemistry Sciences, **5**(2), 23-65 (2020).

[68] A. Mocanu, R. D. Pasca, Gh. Tomoaia, C. Garbo, P. T. Frangopol, O. Horovitz, M. Tomoaia-Cotisel, New procedure to synthesize silver nanoparticles and their interaction with local anesthetics, Int. J. Nanomed., **8**, 3867-3874 (2013).

[69] S. Rapuntean, R. Balint, G. A. Paltinean, Gh. Tomoaia, A. Mocanu, Cs. P. Racz, O. Horovitz, M. Tomoaia-Cotisel, Antibacterial activity of silver nanoparticles obtained by correduction with sodium citrate and tannic acid, Stud. Univ. Babes-Bolyai Chem., **63**(3), 73-85 (2018).

[70] R. Balint, G. A. Paltinean, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, Interaction of silver nanoparticles with vancomycin: an uv-vis study, Stud. Univ. Babes-Bolyai Chem., **64**(2), Tom II, 335-343 (2019).

[71] O. Horovitz, M. Tomoaia-Cotisel, Gh. Tomoaia, L. D. Bobos, O. Cozar, L. Barbu-Tudoran, A. Mocanu, Investigation on the self-assembled arrangement of silver nanoparticles in the presence of protein and amino acids, J. Optoelectron. Adv. Mat.-Symposia, **2**(1), 39-43 (2010).

[72] M. A. Ujica, G. A. Paltinean, A. Mocanu, M. Tomoaia-Cotisel, Silver and Gold Nanoparticles: Challenges and Perspectives, Academy of Romanian Scientists, Annals Series on Biological Sciences, **9**(1), 97-139 (2020).

[73] C. Balan, L.-C. Pop, M. Baia, IR, Raman and SERS analysis of amikacin combined with DFT -based calculations, Spectrochim. Acta A Mol. Biomol. Spectrosc., **214**, 79-85 (2019).

[74] L.-C. Pop, V. Dracopoulos, P. Lianos, Photoelectrocatalytic hydrogen production using nanoparticulate titania and a novel Pt/carbon electrocatalyst: The concept of the "Photoelectrocatalytic Leaf", Appl. Surf. Sci., **333**, 147–151 (2015).

[75] L.-C. Pop, I. Tantis, P. Lianos, Photoelectrocatalytic hydrogen production using nitrogen containing water soluble wastes, Int. J. Hydrog. Energy, **40**, 8304-8310 (2015).

[76] S. Clichici, L. David, B. Moldovan, I. Baldea, D. Olteanu, M. Filip, A. Nagy, V. Luca, C. Crivii, P. Mircea, G. Katona, G. A. Filip, Hepatoprotective effects of silymarin coated gold nanoparticles in experimental cholestasis, Mater. Sci. Eng. C. Mater. Biol. Appl., **115**, 111117 (2020) doi: 10.1016/j.msec.2020.111117.

[77] S. Staroverov, S. Kozlov, A. Fomin, K. Gabalov, A. Volkov, I. Domnitsky, L. Dykman, O. Guliy, Synthesis of silymarin-gold nanoparticle conjugate and analysis of its liver-protecting activity, Curr. Pharm. Biotechnol., **22**(15), 2001-2007 (2021).

[78] A. S. Abdullah, I. E. T. E. Sayed, A. M. A. El-Torgoman, N. A. Alghamdi, S. Ullah, S. Wageh, M. A. Kamel, Preparation and characterization of silymarin-conjugated gold nanoparticles with enhanced anti-fibrotic therapeutic effects against hepatic fibrosis in rats: role of MicroRNAs as molecular targets, Biomedicines, **9**, 1767 (2021). <u>https://doi.org/10.3390/biomedicines9121767</u>.

[79] R. Mohammadinejad, Sh. Pourseyedi, A. Baghizadeh, Sh. Ranjbar, G. A. Mansoori, Synthesis of Silver Nanoparticles Using Silybum Marianum Seed Extract, Int. J. Nanosci. Nanotechnol., **9**(4), 221-226 (2013).

[80] Z. M. Ayad, O. M. Saeed Ibrahim, L. W. Omar, Biosynthesis and characterization of silver nanoparticles by Silybum marianum (silymarin) fruit extract, Adv. Anim. Vet. Sci., **7**(2), 122-130 (2019).

[81] D. Oltean-Dan, G. B. Dogaru, M. Tomoaia-Cotisel, D. Apostu, A. Mester, H. R. C. Benea, M. G. Paiusan, E. M. Jianu, A. Mocanu, R. Balint, C. O. Popa, C. Berce, G. I. Bodizs, A. M. Toader, Gh. Tomoaia, Enhancement of bone consolidation using high-frequency pulsed electromagnetic short-waves and titanium implants coated with biomimetic composite embedded into PLA matrix: in vivo evaluation, Int. J. Nanomed., **14**, 5799-5816 (2019).

[82] D. Oltean-Dan, G.-B. Dogaru, E.-M. Jianu, S. Riga, M. Tomoaia-Cotisel, A. Mocanu, L. Barbu-Tudoran, Gh. Tomoaia, Biomimetic composite coatings for activation of titanium implant surfaces: methodological approach and in vivo enhanced osseointegration, Micromachines, 12(11), 1352 (2021). <u>https://doi.org/10.3390/mi12111352</u>

[83] D. Oltean-Dan, P. T. Frangopol, R. Balint, Gh. Tomoaia, A. Mocanu, M. Tomoaia-Cotisel, Biocompatibility of titanium implants coated with biocomposite in a rat model of femoral fracture, Stud. Univ. Babes-Bolyai Chem., **66**(3), 73-87 (2021).

[84] R. Balint, A. Mocanu, Gh. Tomoaia, S. Riga, M. Tomoaia-Cotisel, Advanced nanomaterials and coated surfaces for orthopedic implants – A review, Annals of the Academy of Romanian Scientists, Series on Physics and Chemistry Sciences, **6**(2), 53-81 (2021).

[85] I. Hodisan, C. Prejmerean, I. Petean, D. Prodan, T. Buruiana, L. Colceriu, L. Barbu-Tudoran, M. Tomoaia-Cotsel, Synthesis and characterization of novel giomers for dental applications, Stud. Univ. Babes-Bolyai Chem., **62**(4), Tom I, 143-154 (2017).

[86] A. Avram, M. Gorea, R. Balint, L. Timis, S. Jitaru, A. Mocanu, M. Tomoaia-Cotisel, Portland cement enriched wth hydroxyapatite for endodontic applications, Stud. Univ. Babes-Bolyai Chem., **62**(4), Tom I, 81-92 (2017).

[87] A. Avram, T. Frentiu, O. Horovitz, A. Mocanu, F. Goga, M. Tomoaia-Cotisel, Hydroxyapatite for removal of heavy metals from wastewater, Stud. Univ. Babes-Bolyai, Chem., 62(4), Tom I, 93-104 (2017).

[88] A. Mocanu, O. Cadar, P. T. Frangopol, I. Petean, Gh. Tomoaia, G. A. Paltinean, Cs. P. Racz, O. Horovitz, M. Tomoaia-Cotisel, Ion release from hydroxyapatite and substituted hydroxyapatites in different immersion liquids: in vitro experiments and theoretical modelling study, R. Soc. Open Sci., **8**(1), 201785, (2021). <u>https://doi.org/10.1098/rsos.201785</u>

[89] C. Garbo, J. Locs, M. D'Este, G. Demazeau, A. Mocanu, C. Roman, O. Horovitz, M. Tomoaia-Cotisel, Advanced Mg, Zn, Sr, Si multi-substituted hydroxyapatites for bone regeneration, Int. J. Nanomed., **15**, 1037-1058 (2020).

[90] S. Rapuntean, P. T. Frangopol, I. Hodisan, Gh. Tomoaia, D. Oltean-Dan, A. Mocanu, C. Prejmerean, O. Soritau, L. Z. Racz, M. Tomoaia-Cotisel, In vitro response of human osteoblasts cultured on strontium substituted hydroxyapatites, Rev. Chim. (Bucharest), **69**(12), 3537-3544 (2018).

[91] O. Cadar, R. Balint, Gh. Tomoaia, D. Florea, I. Petean, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, Behavior of multisubstituted hydroxyapatites in water and simulated body fluid, Stud. Univ. Babes-Bolyai Chem., **62**(4), Tom I, 269-281 (2017).

[92] E. Forizs, F. Goga, A. Avram, A. Mocanu, I. Petean, O. Horovitz, M. Tomoaia-Cotisel, Thermal analysis of pure and multisubstituted hydroxyapatite pastes, Stud. Univ. Babes-Bolyai Chem., **62**(4), Tom I, 173-180 (2017). [93] F. Goga, E. Forizs, G. Borodi, Gh. Tomoaia, A. Avram, R. Balint, A. Mocanu, O. Horovitz,
M. Tomoaia-Cotisel, Behavior of doped hydroxyapatites during the heat treatment, Rev. Chim.
(Bucharest), 68(12), 2907-2913 (2017).

[94] A. Avram, Gh. Tomoaia, A. Mocanu, M. Tomoaia-Cotisel, A review on biomimetic hydroxyapatites for biomedical applications, Academy of Romanian Scientists Annals - Series on Biological Sciences, **9**(2), 106-132 (2020).

[95] A. Mocanu, R. Balint, C. Garbo, L. Timis, I. Petean, O. Horovitz, M. Tomoaia-Cotisel, Low crystallinity nanohydroxyapatite prepared at room temperature, Stud. Univ. Babes-Bolyai Chem., **62**(2), 95-103 (2017).

[96] C. Garbo, M. Sindilaru, A. Carlea, Gh. Tomoaia, V. Almasan, I. Petean, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, Synthesis and structural characterization of novel porous zinc substituted nanohydroxyapatite powders, Particul. Sci. Technol., **35**(1), 29-37 (2017).

[97] P. T. Frangopol, A. Mocanu, V. Almasan, C. Garbo, R. Balint, G. Borodi, I. Bratu, O. Horovitz, M. Tomoaia-Cotisel, Synthesis and structural characterization of strontium substituted hydroxyapatites, Rev. Roum. Chim., **61**(4-5), 337-344 (2016).

[98] A. Mocanu, P. T. Frangopol, R. Balint, O. Cadar, I. M. Vancea, R. Mintau, O. Horovitz, M. Tomoaia-Cotisel, Higuchi model applied to ions release from hydroxyapatites, Stud. Univ. Babes-Bolyai Chem., **66**(3), 195-207 (2021).

[99] F. Goga, E. Forizs, A. Avram, A. Rotaru, A. Lucian, I. Petean, A. Mocanu, M. Tomoaia-Cotisel, Synthesis and thermal treatment of hydroxyapatite doped with magnesium, zinc and silicon, Rev. Chim. (Bucharest), **68**(6), 1193-1200 (2017).

[100] O. Cadar, P. T. Frangopol, Gh. Tomoaia, D. Oltean, G. A. Paltinean, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, Silicon release from hydroxyapatites in water and simulated body fluid, Stud. Univ. Babes-Bolyai Chem., 62(4), Tom I, 67-80 (2017).

[101] G. Furtos, M. Tomoaia-Cotisel, C. Garbo, M. Şenilă, N. Jumate, I. Vida-Simiti, C. Prejmerean, New composite bone cement based on hydroxyapatite and nanosilver, Particul. Sci. Technol., 31(4), 392-398 (2013).

[102] S. A. Kumar, S. Rajeshkumar, S. P. S. Dinesh, A. M. George, R. K. Jain, Antimicrobial activity of silymarin mediated zinc oxide and hydroxy apatite nanoparticles against oral pathogens, Bioinformation, **16**(11), 863-868 (2020).

[103] Z.-S. Tao, X.-J. Wu, M. Yang, H.-G. Xu, Local administration with silymarin could increase osseointegration of hydroxyapatite-coated titanium implants in ovariectomized rats, J. Biomater. Appl., 34(5), 664-672 (2019). <u>https://doi.org/10.1177/0885328219863290</u>.

- [104] A. Di Costanzo, R. Angelico, Formulation strategies for enhancing the bioavailability of silymarin: the state of the art, Molecules, **24**, 2155 (2019). doi:10.3390/molecules24112155
- [105] A. Avram, A. Mocanu, Gh. Tomoaia, M. Tomoaia-Cotisel, Forsterite as an alternative for orthopaedic implants A short review, Annals of the Academy of Romanian Scientists, Series on Physics and Chemistry Sciences, **6**(2), 32-52 (2021).
- [106] M. Gorea, M. A. Naghiu, A. Avram, I. Petean, A. Mocanu, M. Tomoaia-Cotisel, Novel porous forsterite ceramics biocompatibility and bioactivity evaluation, Rev. Chim. (Bucharest), 71(2), 343-351 (2020).
- [107] M. A. Naghiu, M. Gorea, F. Kristaly, M. Tomoaia-Cotisel, A new method for synthesis of forsterite nanomaterials for bioimplants, Ceram.-Silik., **58**(4), 303-307 (2014).
- [108] G. Furtos, M. A. Naghiu, H. Declercq, M. Gorea, C. Prejmerean, O. Pana, M. Tomoaia-Cotisel, Nano forsterite biocomposites for medical applications: Mechanical properties and bioactivity, J. Biomed. Mater. Res. Part B, **104**(7), 1290-1301 (2016).
- [109] M. Gorea, M-A. Naghiu, M. Tomoaia-Cotisel, G. Borodi, Nano and micro-strucure effects on the bioactivity of forsterite powders, Ceram. –Silik., 57(2), 87-91 (2013).
- [110] M. A. Naghiu, M. Gorea, E. Mutch, F. Kristaly, M. Tomoaia-Cotisel, Forsterite nanopowder: structural characterisation and biocompatibility evaluation, J. Mater. Sci. Technol., 29(7), 628-632 (2013).
- [111] M. Gorea, M-A Naghiu, A. Avram, I. Petean, M. Tomoaia-Cotisel, Sintering and characterization of new forsterite ceramics, Stud. Univ. Babes-Bolyai Chem., **64**(2), Tom II, 383-392 (2019).
- [112] A. Avram, M. Gorea, S. Rapuntean, A. Mocanu, G. A. Paltinean, Cs. Varhelyi, Jr., I. Petean, O. Horovitz, M. Tomoaia-Cotisel, In-vitro antibacterial activity of novel nanostructured composites based on forsterite and silver nanoparticles, Rev. Chim. (Bucharest), 71(1), 13-21 (2020).
- [113] A. Avram, S. Rapuntean, M. Gorea, Gh. Tomoaia, A. Mocanu, O. Horovitz, G. Rapuntean,
 M. Tomoaia-Cotisel, In vitro antibacterial effect of forsterite nanopowder: synthesis and characterization, Environ. Sci. Pollut. Res., 29, 77097-77112 (2022).
- [114] G. Furtos, M. A. Naghiu, H. Declercq, M. Gorea, C. Prejmerean, M. Tomoaia-Cotisel, Nano forsterite biocomposites for biomedical application: Mechanical properties and bioactivity, Eur. Cell. Mater., 26(Suppl. 6), 8-8 (2013).

[115] W.-H. Lee, R. Rohanizadeh, C.-Y. Loo, In situ functionalizing calcium phosphate biomaterials with curcumin for the prevention of bacterial biofilm infections, Colloids Surf. B, 206, 111938 (2021). https://doi.org/10.1016/j.colsurfb.2021.111938

[116] Ş. M. Eskitoros-Togay, Y. E. Bulbul, Nursel Dilsiz, Combination of nano-hydroxyapatite and curcumin in a biopolymer blend matrix: Characteristics and drug release performance of fibrous composite material Systems, Int. J. Pharm., **590**, 119933 (2020). https://doi.org/10.1016/j.ijpharm.2020.119933.

[117] W.-H. Lee, C.-Y. Loo, R. Rohanizadeh, Functionalizing the surface of hydroxyapatite drug carrier with carboxylic acid groups to modulate the loading and release of curcumin nanoparticles, Mater. Sci. Eng. C, **99**, 929–939 (2019).

- [118] K. AbouAitah, A. Stefanek, I. M. Higazy, M. Janczewska, A. Swiderska-Sroda, A. Chodara, J. Wojnarowicz, U. Szałaj, S. A. Shahein, A. M. Aboul-Enein, F. Abou-Elella, S. Gierlotka, T. Ciach, W. Lojkowski, Effective targeting of colon cancer cells with piperine natural anticancer prodrug using functionalized clusters of hydroxyapatite nanoparticles, Pharmaceutics, 12, 70, (2020). doi:10.3390/pharmaceutics12010070.
- [119] K. Marycz, A. Smieszek, J. Trynda, P. Sobierajska, S. Targonska, L. Grosman, R. J. Wiglusz, Nanocrystalline hydroxyapatite loaded with resveratrol in colloidal suspension improves viability, metabolic activity and mitochondrial potential in human adipose-derived mesenchymal stromal stem cells (hASCs), Polymers, **11**, 92, (2019). doi:10.3390/polym11010092.
- [120] L. Li, M. Yu, Y. Li, Q. Li, H. Yang, M. Zheng, Y. Han, D. Lu, S. Lu, L. Gui, Synergistic anti-inflammatory and osteogenic n-HA/resveratrol/chitosan composite microspheres for osteoporotic bone regeneration, Bioact. Mater., **6**, 1255–1266 (2021).
- [121] Y. Wu, L. Xia, Y. Zhou, W. Ma, N. Zhang, J. Chang, K. Lin, Y. Xu, X. Jiang, Evaluation of osteogenesis and angiogenesis of icariin loaded on micro/nano hybrid structured hydroxyapatite granules as a local drug delivery system for femoral defect repair, J. Mater. Chem. B, 3(24), 4871-4883 (2015).
- [122] J. Fan, L. Bi, T. Wu, L. Cao, D. Wang, K. Nan, J. Chen, D. Jin, S. Jiang, G. Pei, A combined chitosan/nano-size hydroxyapatite system for the controlled release of icariin, J. Mater. Sci. Mater. Med., 23, 399–407 (2012).
- [123] S. Kongsri, P. L. Na Ayuttaya, S. Yookhum, S. Techawongstein, S. Chanthai, Characterization of hydroxyapatite nanoparticles from fish scale waste and its adsorption of carotenoids, Asian J. Chem., **25**(10), 5847-5850 (2013).

- [124] S. Kongsri, S. Techawongstien, S. Chanthai, The Synergistic effect of anionic surfactant on adsorption enhancement of the carotenoids extract using mesoporous hydroxyapatite nanoparticles, Orient. J. Chem., **31**(3), 1331-1343 (2015).
- [125] M. Tomoaia-Cotisel, P. J. Quinn, *Biophysical properties of carotenoids*, in book Subcellular Biochemistry, Fat-Soluble Vitamins, Editors: P. J. Quinn and V. E. Kagan, (Plenum Press, New York, USA, 1998), vol. **30**, chapter 10, pp.219-242.
- [126] G. Britton, Carotenoid research: History and new perspectives for chemistry in biological systems, BBA Mol. Cell Biol. Lipids, 1865, 158699 (2020). https://doi.org/10.1016/j.bbalip.2020.158699
- [127] M. Tomoaia, A. Ioanette, E. Chifu, *Behavior of β-apo-8'-carotenal at the oil/water and air/water interfaces*. In Proceedings of the International Conference of Colloid and Surface Science, E. Wolfram, ed., (Akadémiai Kiadó, Budapest, Ungaria, 1975), Vol. 1, pp. 559-566.
- [128] F. Nabi, M. A. Arain, N. Rajput, M. Alagawany, J. Soomro, M. Umer, F. Soomro, Z. Wang,
 R. Ye, J. Liu, Health benefits of carotenoids and potential application in poultry industry: A review, J. Anim. Physyol. Anim. Nutr., **104**(6), 1809-1818 (2020).
- [129] O. Horovitz, Gh. Tomoaia, Cs. Racz, A. Mocanu, L.-D. Bobos, M. Tomoaia-Cotisel, Surface properties of some carotenoids spread in monolayers at the air/water interface. Experimental and computational approach, Cent. Eur. J. Chem., **4**(3), 489–501 (2006).
- [130] E. J. Johnson, The role of carotenoids in human health, Nutr. Clin. Care, 5(2), 56-65 (2002).
- [131] A. F. G. Cicero, A. Colletti, Effects of carotenoids on health: are all the same? results from clinical trials, Curr. Pharm. Des., **23**, 2422-2427 (2017).
- [132] E. Chifu, J. Zsakó, M. Tomoaia-Cotisel, Xanthophyll films: I. Single-component monolayers at the air/water interface, J. Colloid Interface Sci., 95(2), 346-354 (1983).
- [133] M. Tomoaia-Cotisel, E. Chifu, Xanthophyll films: II. Two-component monolayers of some xanthophylls and egg lecithin at the air/water interface, J. Colloid Interface Sci., **95**(2), 355-361 (1983).
- [134] M. Tomoaia-Cotisel, E. Chifu, J. Zsakó, Xanthophyll films: III. Two-component monolayers of some xanthophylls and L-α-dipalmitoyl lecithin at the air/water interface, Stud. Univ. Babeş-Bolyai Chem., **31**(2), 80-89 (1986).
- [135] E. Chifu, M. Tomoaia, A. Ioanette, Behavior of canthaxanthin at the benzene-water and airwater interfaces, Gazz. Chim. Ital., **105**(11), 1225-1232 (1975).
- [136] E. Chifu, M. Tomoaia-Cotişel, Z. Andrei, Mixed monolayers of canthaxanthin with lipids, Stud. Univ. Babeş-Bolyai Chem., 24(2) 63-67 (1979).

[137] M. E. Orczyk, J. Samoc, M. Swiatkiewicz, N. Manickam, M. Tomoaia-Cotişel, P. N. Prasad, Optical heterodyning of the phase-tuned femtosecond optical Kerr gate signal for the determination of complex third-order susceptibilities, Appl. Phys. Lett., 60(23), 2837-2839 (1992).

- [138] M. Mirahmadi, S. Azimi-Hashemi, E. Saburi, H. Kamali, M. Pishbin, F. Hadizadeh, Potential inhibitory effect of lycopene on prostate cancer, Biomed. Pharmacother., **129**, 110459 (2020). <u>https://doi.org/10.1016/j.biopha.2020.110459</u>
- [139] N. Y. Lee, Y. Kim, Y. S. Kim, J.-H. Shin, L. P. Rubin, Y. Kim, β-Carotene exerts anticolon cancer effects by regulating M2 macrophages and activated fibroblasts, J. Nutr. Biochem., 82, 108402 (2020). doi: 10.1016/j.jnutbio.2020.108402
- [140] P. Joos, A. Tomoaia-Cotisel, A. J. Sellers, M. Tomoaia-Cotisel, Adsorption kinetics of some carotenoids at the oil/water interface, Colloids Surf. B, 37(3-4), 83-91 (2004).
- [141] M. Tomoaia-Cotisel, J. Zsakó, E. Chifu, D. A. Cadenhead, Relaxation phenomena in apocarotenoid monolayers, Langmuir, **6**(1), 191-197 (1990).
- [142] Gh. Tomoaia, A. Tomoaia-Cotisel, M. Tomoaia-Cotisel, A. Mocanu, Kinetic study of adsorption of some biocompounds at the oil/water interface, Centr. Eur. J. Chem., **3**(2), 347-360 (2005).
- [143] M. Tomoaia-Cotisel, J. Zsakó, E. Chifu, P. J. Quinn, Intermolecular interactions in lipid carotenoid monolayers, Biochem. J., **248**(3), 877-882 (1987).
- [144] M. Tomoaia-Cotisel, E. Chifu, J. Zsakó, Mixed monolayers of egg lecithin and carotenoids, Colloids Surf., 14(3-2), 239-246 (1985).
- [145] E. Chifu, M. Tomoaia-Cotisel, Mixed films of carotenoid pigments and lipids at the air/water interface, Rev. Chim. (Bucharest), **33**(2), 125-131 (1982).
- [146] E. Chifu, M. Tomoaia-Cotisel, Insoluble monolayers of lecithin and carotenoid pigments, Rev. Chim. (Bucharest), 24(7), 979-986 (1979).
- [147] E. Chifu, M. Tomoaia, E. Nicoară, A. Olteanu, Isozeaxanthin films at the oil/water and air/water interfaces, Rev. Roum. Chim., **23**(8), 1163-1169 (1978).
- [148] J. Zsakó, E. Chifu, M. Tomoaia-Cotisel, Rotating rigid-plate model of carotenoid molecules and the behavior of their monolayers at the air/water interface, Gazz. Chim. Ital., **109**(11-12), 663-668 (1979).
- [149] M. Tomoaia-Cotisel, E. Chifu, Mixed insoluble monolayers with β -apo-8'-carotenoic acid ethyl ester, Gazz. Chim. Ital., **109**(6-7), 371-375 (1979).
- [150] E. Chifu, M. Tomoaia-Cotişel, A. Ioanette, Mixed insoluble monolayers of cholesterol and β -apo-8'-carotenal, Gazz. Chim. Ital., **109**(6-7), 397-398 (1979).

- [151] M. Tomoaia-Cotisel, E. Chifu, V. Tămaş, V. Mărculeţiu, Behavior of some apocarotenoid derivatives at the air/water interface, Rev. Roum. Chim., 25(2), 175-180 (1980).
- [152] M. Tomoaia-Cotisel, E. Chifu, Carotenoid pigment films at fluid interfaces, Rev. Chim. (Bucharest), 32(11), 1063-1069 (1981).
- [153] E. Chifu, M. Tomoaia-Cotisel, Carotene and protide films at the oil/water interface, Stud. Univ. Babes-Bolyai Chem., 26(2), 3-8 (1981).
- [154] I. Cojocaru, A. Tomoaia-Cotisel, A. Mocanu, T. Yupsanis, M. Tomoaia-Cotisel, The effects of globular protein from aleurone cells of barley on stearic acid monolayers, Rev. Chim. (Bucharest), 68(7), 1470-1475 (2017).
- [155] J. Zsako, M. Tomoaia-Cotișel, E. Chifu, Insoluble mixed monolayers: I. Phase equilibria at the collapse of binary monolayers at gas/liquid interfaces, J. Colloid Interface Sci., **102**(1), 186-205 (1984).
- [156] R. D. Pasca, Gh. Tomoaia, A. Mocanu, I. Petean, A. G. Paltinean, O. Soritau, M. Tomoaia-Cotisel, Porous collagen scaffolds for bone regeneration, Stud. Univ. Babes-Bolyai Chem., 60(3), 257-264 (2015).
- [157] Gh. Tomoaia, O. Soritau, M. Tomoaia-Cotisel, L. B. Pop, A. Pop, A. Mocanu, O. Horovitz,
 L. D. Bobos, Scaffolds made of nanostructured phosphates, collagen and chitosan for cell culture,
 Powder. Technol., 238, 99-107 (2013).
- [158] Gh. Tomoaia, M. Tomoaia-Cotisel, L.-B. Pop, A. Pop, O. Horovitz, A. Mocanu, N. Jumate, L.-D. Bobos, Synthesis and characterization of some composites based on nanostructured phosphates, collagen and chitosan, Rev. Roum. Chim., 56(10-11), 1039-1046 (2011).
- [159] M. Tomoaia-Cotisel, J. Zsakó, A. Mocanu, E. Chifu, P. J. Quinn, Monolayer properties of membrane lipids of the extreme halophile Halobacterium cutirubrum at the air/water interface, BBA-Biomembranes, 942(2), 295-304 (1988).
- [160] M. Tomoaia-Cotisel, E. Chifu, J. Zsakó, A. Mocanu, P. J. Quinn and M. Kates, Monolayer properties of archaeol and caldarchaeol polar lipids of a methanogenic archaebacterium, Methanospirillum hungatei, at the air/water interface, Chem. Phys. Lipids, 63(1-2), 131-138 (1992).
- [161] J. Zsakó, M. Tomoaia-Cotisel, E. Chifu, A. Mocanu, P. T. Frangopol, Procaine interactions with phospholipid monolayers at the air/water interface, Gazz. Chim. Ital., **124**(1), 5-9 (1994).
- [162] M. Tomoaia-Cotisel, A. Mocanu, Phase transitions in phospholipid monolayers studied by atomic force microscopy and Langmuir-Blodgett technique, Rev. Chim. (Bucharest), 59(11), 1230-1233 (2008).

- [163] M. Tomoaia-Cotisel, I. W. Levin, Thermodynamic study of the effects of ursodeoxycholic acid and ursodeoxycholate on aqueous dipalmitoyl phosphatidylcholine bilayer dispersions, J. Phys. Chem. B, **101**(42), 8477-8485 (1997).
- [164] M. Tomoaia-Cotisel, J. Zsakó, E. Chifu, P. J. Quinn, Influence of electrolytes on the monolayers properties of saturated galactolipids at the air/water interface, Chem. Phys. Lipids, 34(1), 55-64 (1983).
- [165] M. Tomoaia-Cotisel, A. Sen, P. J. Quinn, Surface-active properties of 1, 2distearoylgalactosylglycerols, J. Colloid Interface Sci., 94(2), 390-398 (1983).
- [166] Cs-P. Racz, Sz. Santa, M. Tomoaia-Cotisel, Gh. Borodi, I. Kacso, A. Pirnau, I. Bratu, Inclusion of α -lipoic acid in β -cyclodextrin. Physical-chemical and structural characterization, J. Incl. Phenom. Macrocycl. Chem., **76**(1-2), 193-199 (2013).
- [167] Cs. P. Racz, R. D. Pasca, S. Santa, I. Kacso, G. Tomoaia, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, Inclusion complex of beta-cyclodextrin and quercetin. Thermodynamic approach, Rev. Chim. (Bucharest), 62(10), 992-997 (2011).
- [168] Cs. P. Rácz, G. Borodi, M. M. Pop, I. Kacso, S. Santa, M. Tomoaia-Cotisel, Structure of the inclusion complex of-cyclodextrin with lipoic acid from laboratory powder diffraction data, Acta Crystallogr. B, 68(2), 164-170 (2012).
- [169] C. G. Floare, M. Bogdan, M. Tomoaia-Cotişel, A. Mocanu, 1H NMR spectroscopic characterization of inclusion complex of desferrioxamine B chelator and β-cyclodextrin, J. Mol. Struct., **1248**, 131477 (2022). <u>https://doi.org/10.1016/j.molstruc.2021.131477</u>
- [170] M. Tomoaia-Cotisel, J. Zsakó, A. Mocanu, I. Albu, E. Chifu, Relaxation phenomena in fatty acid monolayers, Stud. Univ. Babes-Bolyai Chem., **32**(1), 58-67 (1987).
- [171] M. Tomoaia-Cotisel, J. Zsako, A. Mocanu, M. Lupea, E. Chifu, Insoluble mixed monolayers: III. The ionization characteristics of some fatty acids at the air/water interface, J. Colloid Interface Sci., 117(2), 464-476 (1987).
- [172] M. Tomoaia-Cotisel, J. Zsakó, E. Chifu, State equations of fatty acid monolayers, Stud. Univ. Babes-Bolyai Chem., 33(2), 54-60 (1988).
- [173] E. Chifu, J. Zsakó, M. Tomoaia-Cotisel, A. Mocanu, A comparative study of some fatty acid monolayers at the air/water interface, Stud. Univ. Babes-Bolyai Chem., **34**(2), 3-9 (1989).
- [174] M. Tomoaia-Cotisel, *The nanostructure formation of the globular seed storage protein on different solid surfaces studied by atomic force microscopy*, in Convergence of Micro-NanoBiotechnologies, Series in Micro and Nanoengineering, Eds: Maria Zaharescu, Emil Burzo,

Lucia Dumitru, Irina Kleps and Dan Dascalu, (Romanian Academy Press, Bucharest, Romania, 2006), Vol. 9, pp. 147-161.

- [175] M. Tomoaia-Cotisel, A. Tomoaia-Cotisel, T. Yupsanis, Gh. Tomoaia, I. Balea, A. Mocanu, Cs. Racz, Coating layers of major storage protein from aleurone cells of barley studied by atomic force microscopy, Rev. Roum. Chim., **51**(12), 1181-1185 (2006).
- [176] M. Tomoaia-Cotisel, R. D. Pasca, O Horovitz, A. Mocanu, Surface potentials of cholesterol and dimyristoyl phosphatidylcholine monolayers at the air/water interface, Rev. Roum. Chim., 56(10-11), 1047-1053 (2011).
- [177] A. Mocanu, P.J. Quinn, C. Nicula, S. Riga, Gh. Tomoaia, C.-A. Bardas, M. Tomoaia-Cotisel, Interaction of local anesthetic procaine with phospholipid model membrane, Rev. Roum. Chim., 66(10-11), 855-869 (2021). <u>https://doi.org/10.33224/rrch.2021.66.10-11.09</u>
- [178] M. Tomoaia-Cotisel, J. Zsako, E. Chifu, Dipalmitoyl lecithin and egg lecithin monolayers at an air/water interface, Ann. Chim. (Rome), **71**(3-4), 189-200 (1981).
- [179] M. Tomoaia-Cotisel, D. V. Pop-Toader, U. V. Zdrenghea, Gh. Tomoaia, O. Horovitz, A. Mocanu, Desferal effect on human erythrocyte membrane. An atomic force microscopy analysis, Stud. Univ. Babes-Bolyai Chem., 54(4), 285-296 (2009).
- [180] B. Asgharian, D. A. Cadenhead, M. Tomoaia-Cotisel, An epifluorescent microscopy study of the effects of procaine on model membrane systems, Langmuir, **9**(1), 228-232 (1993).
- [181] U. V. Zdrenghea, Gh. Tomoaia, D.-V. Pop-Toader, A. Mocanu, O. Horovitz, M. Tomoaia-Cotisel, Procaine effect on human erythrocyte membrane explored by atomic force microscopy, Comb. Chem. High Throughput Screen., **14**(4), 237-247 (2011).
- [182] C. Tan, J. Xue, X. Lou, S. Abbas, Y. Guan, B. Feng, X. Zhanga, S. Xia, Liposomes as delivery systems for carotenoids: comparative studies of loading ability, storage stability and in vitro release, Food Funct., **5**(6), 1232-1240 (2014).
- [183] W. K. Subczynski, A. Wisniewska, J. Widomska, Location of macular xanthophylls in the most vulnerable regions of photoreceptor outer-segment membranes, Arch. Biochem. Biophys., 504, 61-66 (2010).
- [184] J. Widomska, J. P. SanGiovanni, W. K. Subczynski, Why Is Zeaxanthin the most concentrated xanthophyll in the central fovea?, Nutrients, **12**(5), 1333 (2020) <u>https://doi.org/10.3390/nu12051333</u>
- [185] E. H. Papaioannou, M. Liakopoulou –Kyriakides, A. J. Karabelas, Natural origin lycopene and its 'green' downstream processing, Crit. Rev. Food Sci. Nutr., **56**(4), 686-709 (2015).

- [186] M. Bacanli, N. Başaran, A. A. Başaran, Lycopene: is it beneficial to human health as an antioxidant?, Turk. J. Pharm. Sci., **14**(3), 311-318 (2017).
- [187] M. Camara, M. de Cortes Sanchez-Mata, V. Fernandez-Ruiz, R. M. Camara, S. Manzoor, J.
 O. Caceres, *Lycopene: A Review of Chemical and Biological Activity Related to Beneficial Health Effects*, in book Studies in Natural Products Chemistry, Atta-ur-Rahman Eds., (Academic Press, Elsevier, Oxford, UK, 2013), Chapter 11, Vol. 40, pp.383-426.
- [188] N. Ford, J. W. Erdman Jr., *Lycopene and Cancer*, in book Carotenoids and Human Health, Nutrition and Health, S.A. Tanumihardjo (ed.), (Humana Press, New York, USA, 2013), Section II, chapter 12, pp. 193-214
- [189] W. I. Gruszecki, K. Strzałka, Carotenoids as modulators of lipid membrane physical properties, Biochim. Biophys. Acta Mol. Basis Dis., **1740**(2), 108-115 (2005).
- [190] R. Rivera-Madrid, V. M. Carballo-Uicab, Y. Cárdenas-Conejo, M. Aguilar-Espinosa, R. Siva, *Overview of carotenoids and beneficial effects on human health*, in book Carotenoids: Properties, Processing and Applications, edited by C. M.Galanakis, (Academic Press, Oxford, UK, 2019), chapter 1, pp. 1-40.
- [191] L. Bogacz-Radomska, J. Harasym, β -Carotene—properties and production methods, Food Qual. Saf., **2**(2), 69–74 (2018).
- [192] R. Pritwani, P. Mathur, β -carotene content of some commonly consumed vegetables and fruits available in Delhi, India, J. Nutr. Food Sci., **7**, 5 (2017).
- [193] S. Jeyakodi, A. Krishnakumar, D. K. Chellappan, Beta Carotene -therapeutic potential and strategies to enhance its bioavailability, Nutri. Food Sci. Int. J., 7(4), 555716 (2018). DOI: 10.19080/NFSIJ.2018.07.555716
- [194] R. Vishwanathan, E. J. Johnson, *Lutein and Zeaxanthin and Eye Disease*, in book Carotenoids and Human Health, Nutrition and Health, S.A. Tanumihardjo (ed.), (Humana Press, New York, USA, 2013), Section II, chapter 13, 215-235.
- [195] J. W. Smith, R. B. Rogers, S. Jeon, S. S. Rubakhin, L. Wang, J. V. Sweedler, M. Neuringer, M. J. Kuchan, J. W. Erdman, Jr, Carrot solution culture bioproduction of uniformly labeled 13Clutein and in vivo dosing in non-human primates, Exp. Biol. Med. (Maywood)., 242(3), 305–315 (2017).
- [196] X. Gong, J. R. Smith, H. M. Swanson, L. P. Rubin, Carotenoid lutein selectively inhibits breast cancer cell growth and potentiates the effect of chemotherapeutic agents through ROSmediated mechanisms, Molecules, 23, 905 (2018). doi:10.3390/molecules23040905

- [197] M. O. Becerra, L. M. Contreras, M. Hsieh Lo, J. M. Díaz, G. C. Herrera, Lutein as a functional food ingredient: Stability and bioavailability, J. Func. Foods, 66, 103771 (2020). https://doi.org/10.1016/j.jff.2019.103771
- [198] R. R. Ambati, P. S. Moi,S. Ravi, R. G. Aswathanarayana, Astaxanthin: sources, extraction, stability, biological activities and its commercial applications—A Review, Mar. Drugs., 12(1), 128–152 (2014).
- [199] E. Yamashita, Astaxanthin as a Medical Food, Funct. Food Health Dis., **3**(7), 254-258 (2013).
- [200] P. Kidd, Astaxanthin, cell membrane nutrient with diverse clinical benefits and anti-aging potential, Altern. Med. Rev., **16**(4), 355-364 (2011).
- [201] S. Chintong, W. Phatvej, U. Rerk-Am, Y. Waiprib, W. Klaypradit, In Vitro antioxidant, antityrosinase, and cytotoxic activities of astaxanthin from shrimp waste, Antioxidants, 8(5), 128 (2019). doi:10.3390/antiox8050128
- [202] S.-Q. Sun, Y.-X. Zhao, S.-Y. Li, J.-W. Qiang, Y.-Z. Ji, Anti-Tumor effects of astaxanthin by inhibition of the expression of STAT3 in prostate cancer, Mar. Drugs, 18(8), 415 (2020). doi: 10.3390/md18080415
- [203] S. Tsuji, S. Nakamura, T. Maoka, T. Yamada, T. Imai, T. Ohba, T. Yako, M. Hayashi, K. Endo, M. Saio, H. Hara, M. Shimazawa, Antitumour effects of astaxanthin and adonixanthin on glioblastoma, Mar. Drugs, 18(9), 474 (2020) doi: 10.3390/md18090474
- [204] J. Ávila-Román, S. García-Gil, A. Rodríguez-Luna, V. Motilva, E. Talero, Anti-Inflammatory and anticancer effects of microalgal carotenoids, Mar. Drugs, **19**(10), 531 (2021) doi: 10.3390/md19100531.
- [205] A. G. Murillo, S. Hu, M. L. Fernandez, Zeaxanthin: metabolism, properties, and antioxidant protection of eyes, heart, liver, and skin, Antioxidants (Basel), **8**(9), 390 (2019) doi: 10.3390/antiox8090390.
- [206] A. Sujak, J. Gabrielska, J. Milanowska, P. Mazurek, K. Strzaakae, W. I. Gruszecki, Studies on canthaxanthin in lipid membranes, Biochim. Biophys. Acta, 1712, 17 – 28 (2005).
- [207] B. A. Rebelo, S. Farrona, M. R. Ventura, R. Abranches, Canthaxanthin, a red-hot carotenoid: applications, synthesis, and biosynthetic evolution, Plants, 9, 1039 (2020). doi:10.3390/plants9081039
- [208] P. Palozza, N. Maggiano, G. Calviello, P. Lanza, E. Piccioni, F. O. Ranelletti, G. M. Bartoli, Canthaxanthin induces apoptosis in human cancer cell lines, Carcinogenesis, 19(2) 373–376 (1998).

- [209] L. Garavaglia, S. Gallettia, D. Tedesco, Silymarin and lycopene administration in periparturient dairy cows: effects on milk production and oxidative status, N. Z. Vet. J., **63**(6), 313-318 (2015).
- [210] E. Yurtcu, E. Kasapoğlu, F. İ. Şahin, Protective eff ects of β -carotene and silymarin on human lymphocytes, Turk. J. Biol., **36**, 47-52 (2012).
- [211] R. P. Assis, C. A. Arcaro, V. O. Gutierres, J. O. Oliveira, P. I. Costa, A. M. Baviera, I. L. Brunetti, Combined effects of curcumin and lycopene or bixin in yoghurt on inhibition of LDL oxidation and increases in HDL and Paraoxonase levels in streptozotocin-diabetic rats, Int. J. Mol. Sci., 18, 332 (2017). doi:10.3390/ijms18040332
- [212] N. Rajput, M. Naeem, S. Ali, J. F. Zhang, L. Zhang, T. Wang, The effect of dietary supplementation with the natural carotenoids curcumin and lutein on broiler pigmentation and immunity, Poultry Science, **92**(5), 1177-1185 (2013).
- [213] B. M. Steiner, V. Shukla, D. J. McClements, Y. O. Li, M. Sancho-Madriz, G. Davidov-Pardo, Encapsulation of lutein in nanoemulsions stabilized by resveratrol and maillard conjugates, J. Food Sci., 84(9), 2421-2431 (2019).
- [214] A. Kawamura, W. Aoi, R. Abe, Y. Kobayashi, M. Kuwahata, A. Higashi, Astaxanthin-, β carotene-, and resveratrol-rich foods support resistance training-induced adaptation, Antioxidants (Basel)., **10**(1), 113 (2021) doi: 10.3390/antiox10010113.
- [215] A. Kawamura, W. Aoi, R. Abe, Y. Kobayashi, S. Wada, M. Kuwahata, A. Higashi, Combined intake of astaxanthin, b-carotene, and resveratrol elevates protein synthesis during muscle hypertrophy in mice, Nutrition, 69, 110561 (2020) <u>https://doi.org/10.1016/j.nut.2019.110561</u>.
- [216] L. C. Pop, M. Baibarac, I. Anghel, L. Baia, Gypsum composite boards incorporating phase change materials: A review, J. Nanosci. Nanotechnol., **21**, 2269–2277 (2021).